

THE IMPACT OF CLINICAL PHARMACOLOGY IN HIV CURE RESEARCH


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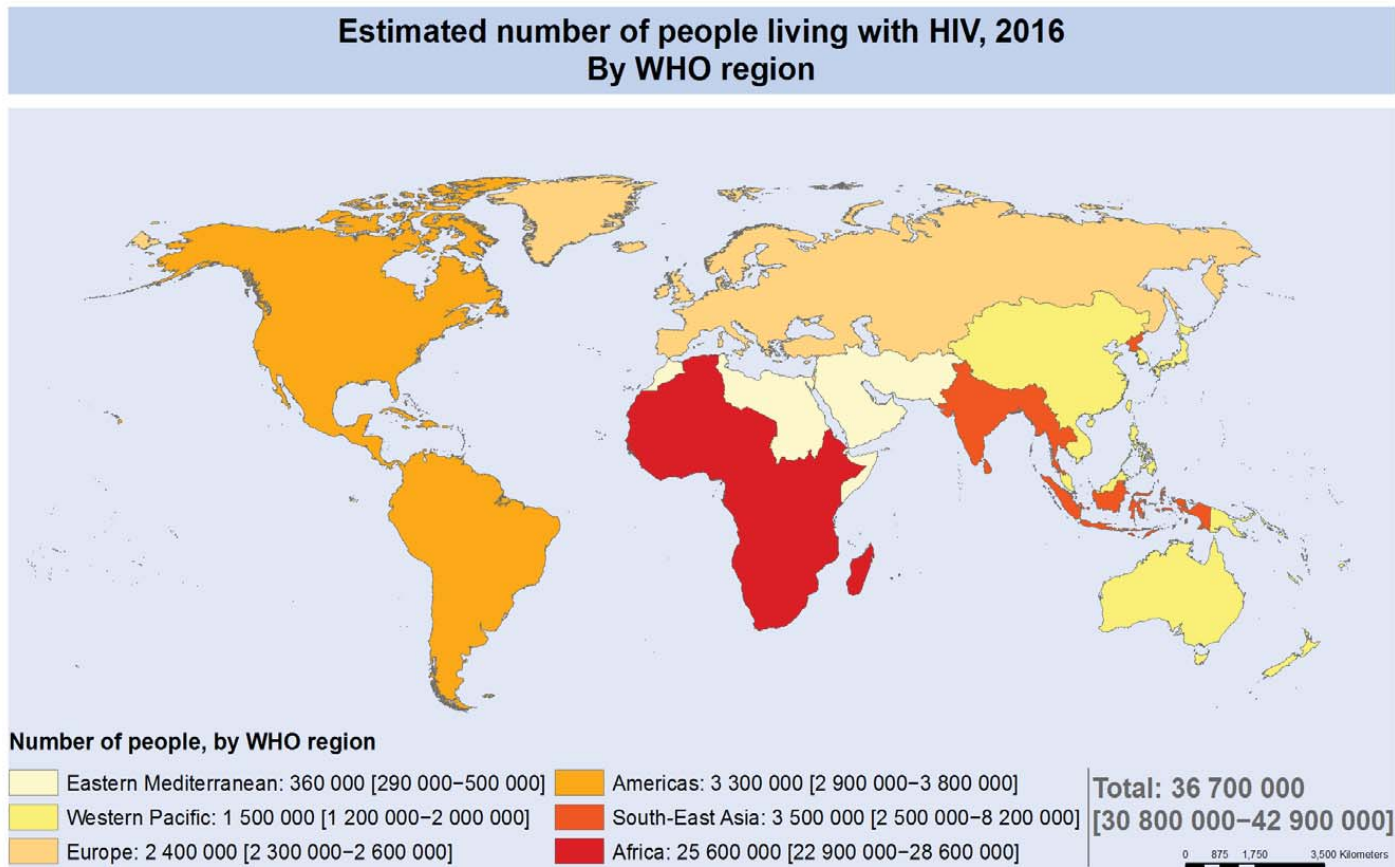
UNC Eshelman School of Pharmacy

QUESTIONS

- ▶ Why do we need a cure for HIV?
 - ▶ Why don't we already have a cure?
 - ▶ What are current strategies for cure?
 - ▶ What challenges need pharmacology insights?
- 
- A decorative graphic consisting of several parallel white lines of varying lengths, slanted diagonally from the bottom right towards the top right, set against the dark blue background.

ONE REASON WE NEED A CURE: GLOBAL HIV BURDEN 2016

People living with HIV	36.7 million
New HIV infections	1.8 million
AIDS-related deaths	1.0 million
5,000 new infections per day	
64% sub-Saharan Africa	
43% female adults	



The boundaries and names shown and the designations used on this map do not imply the expression of any opinion whatsoever on the part of the World Health Organization concerning the legal status of any country, territory, city or area or of its authorities, or concerning the delimitation of its frontiers or boundaries. Dotted and dashed lines on maps represent approximate border lines for which there may not yet be full agreement.

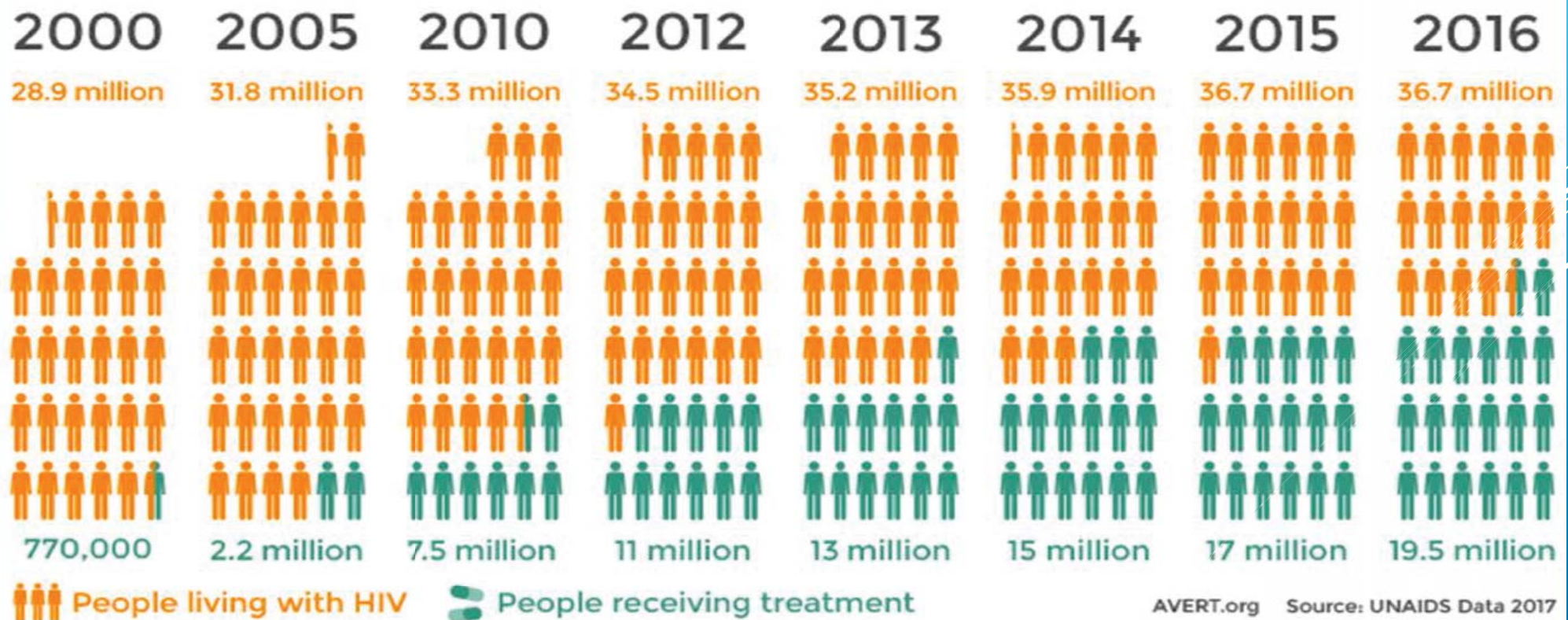
Data Source: World Health Organization
Map Production: Information Evidence and Research (IER)
World Health Organization



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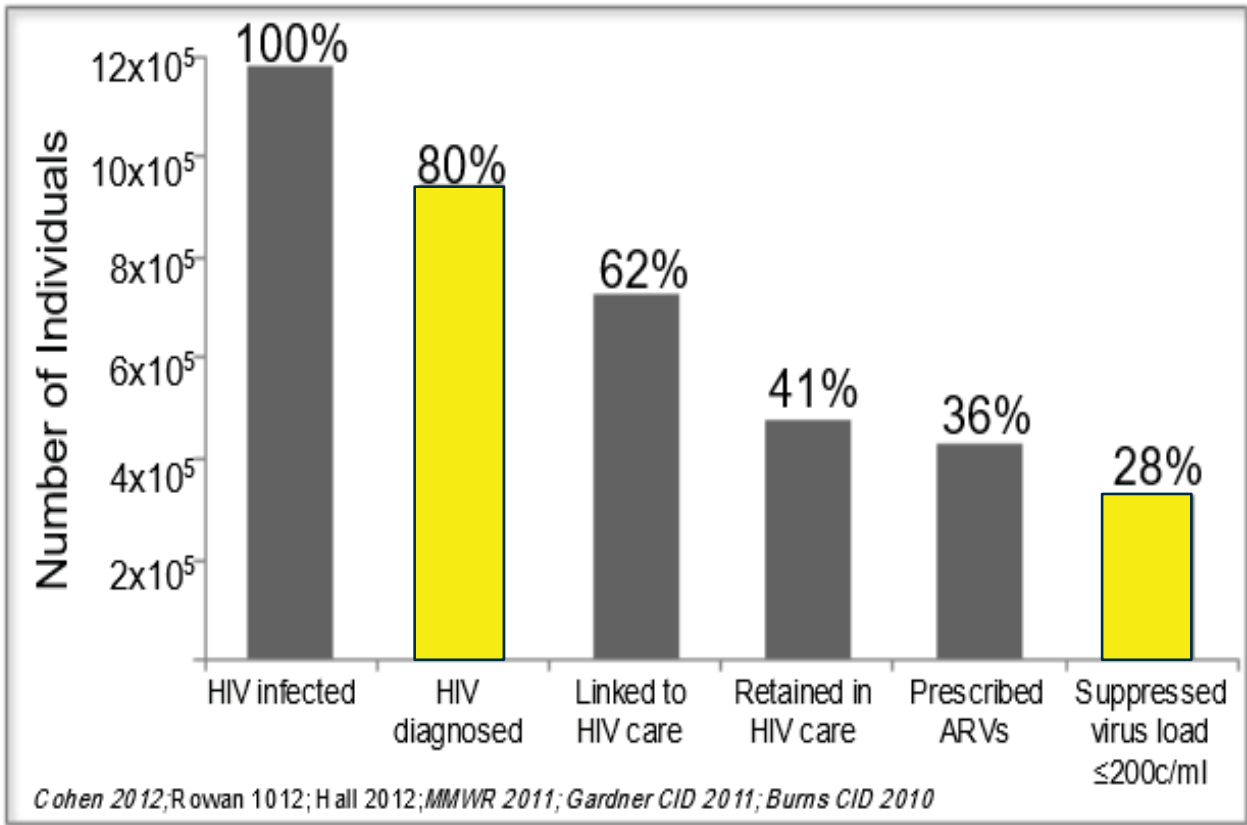
INCREASING ACCESS TO TREATMENT

Number of people living with HIV and accessing treatment globally

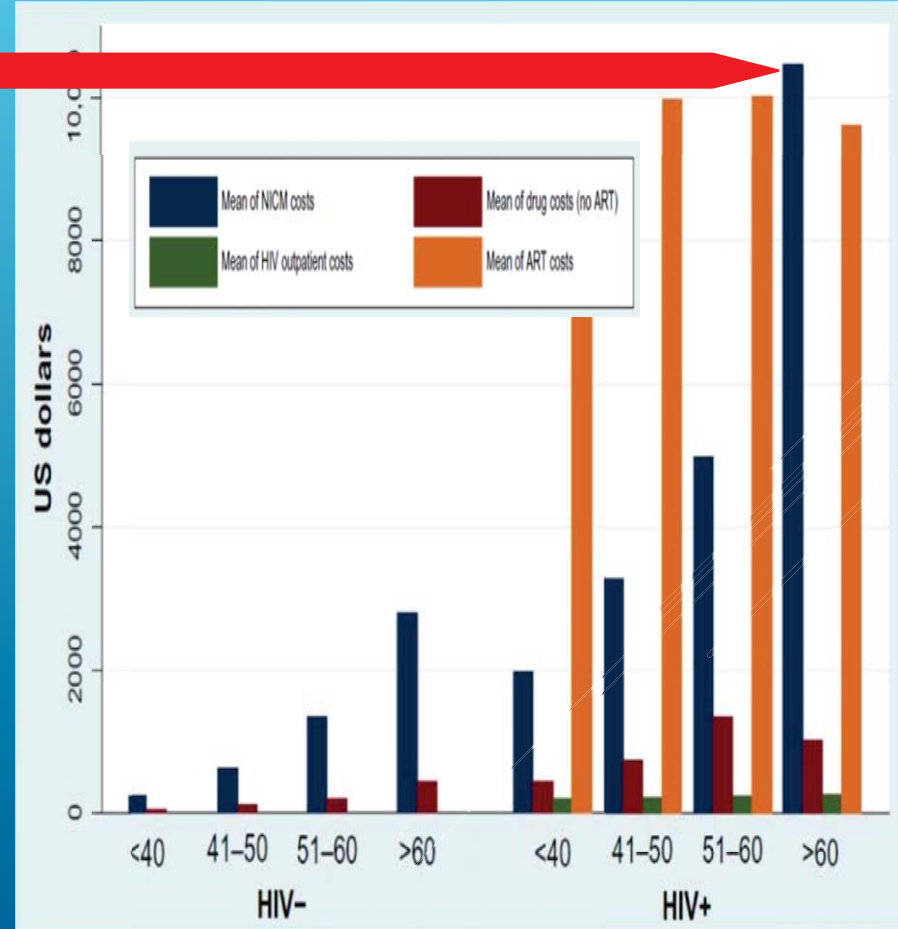
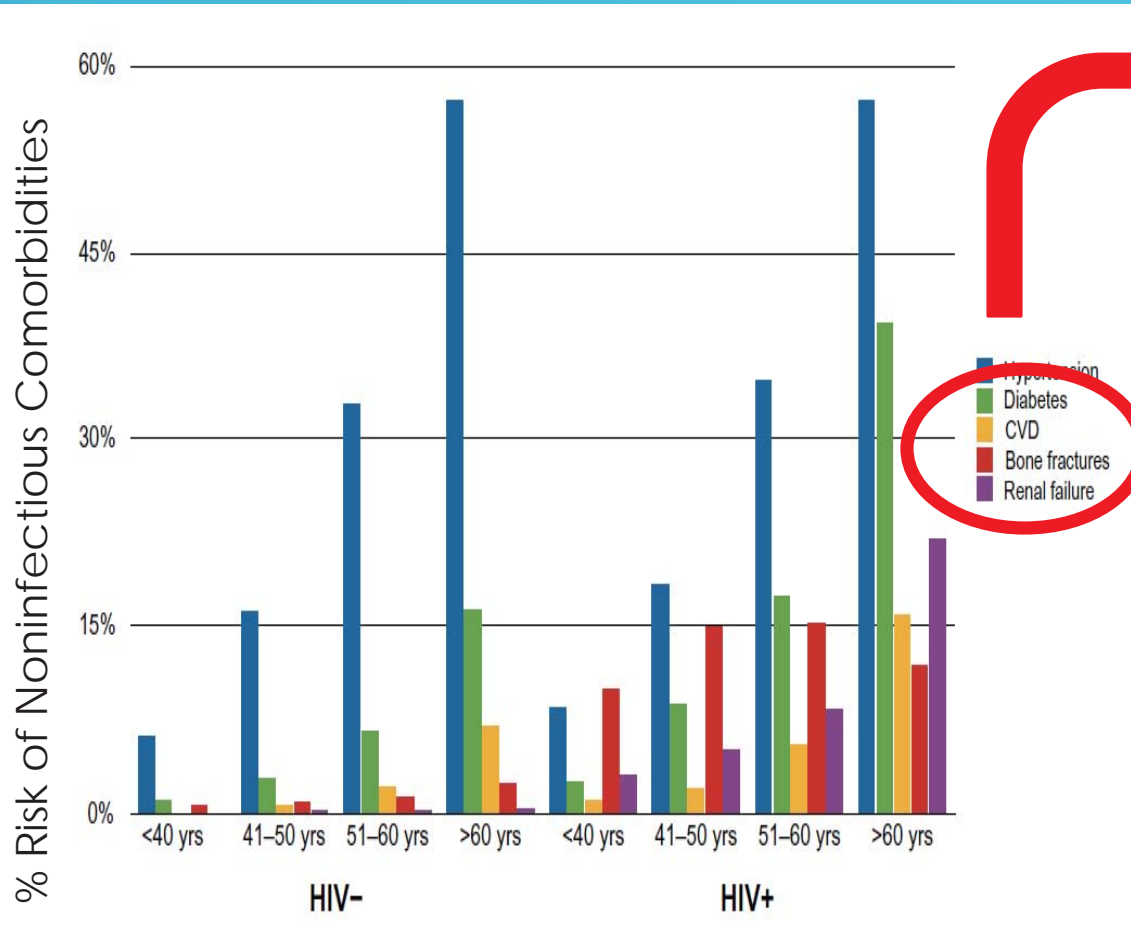


ANOTHER REASON WE NEED A CURE: THE TREATMENT CASCADE

Number and percentage of HIV-infected persons engaged in selected stages of the continuum of HIV care in the United States



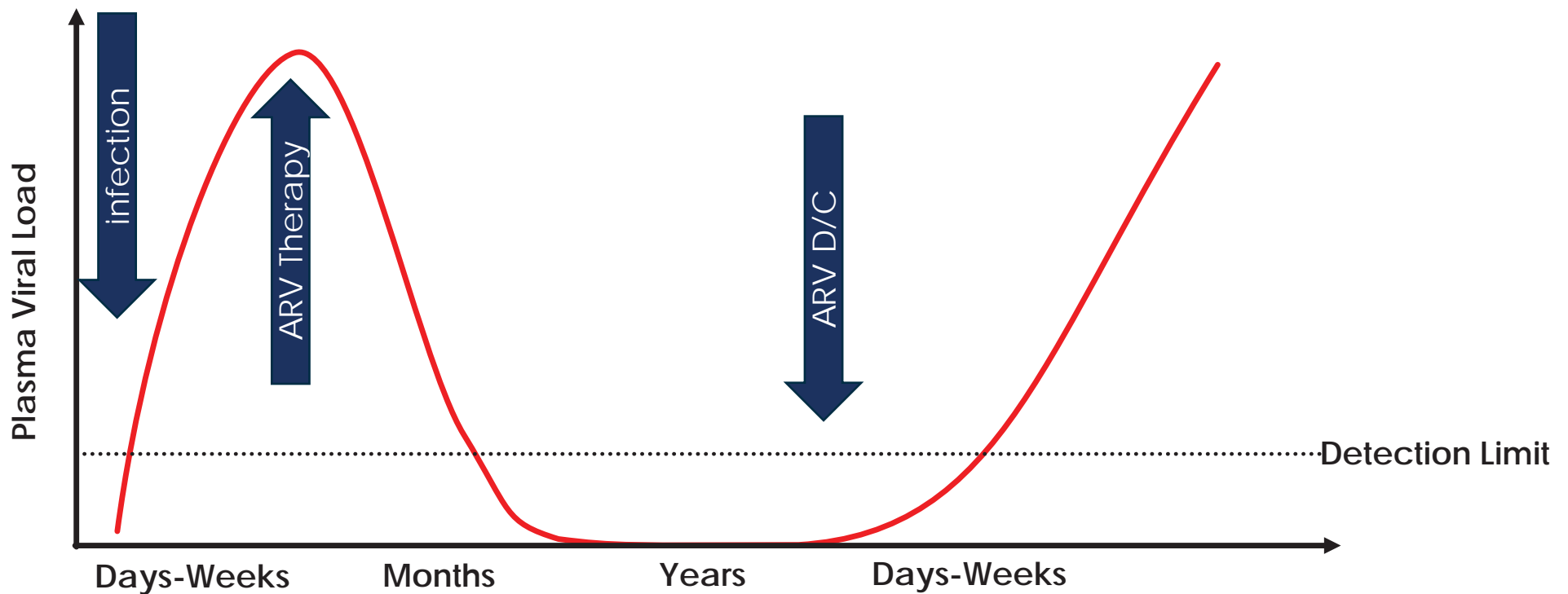
ANOTHER REASON WE NEED A CURE: INCREASED COMORBIDITIES (ARVS AND INFLAMMATION)



WHY DON'T WE HAVE A CURE?

HIV PERSISTENCE ON TREATMENT

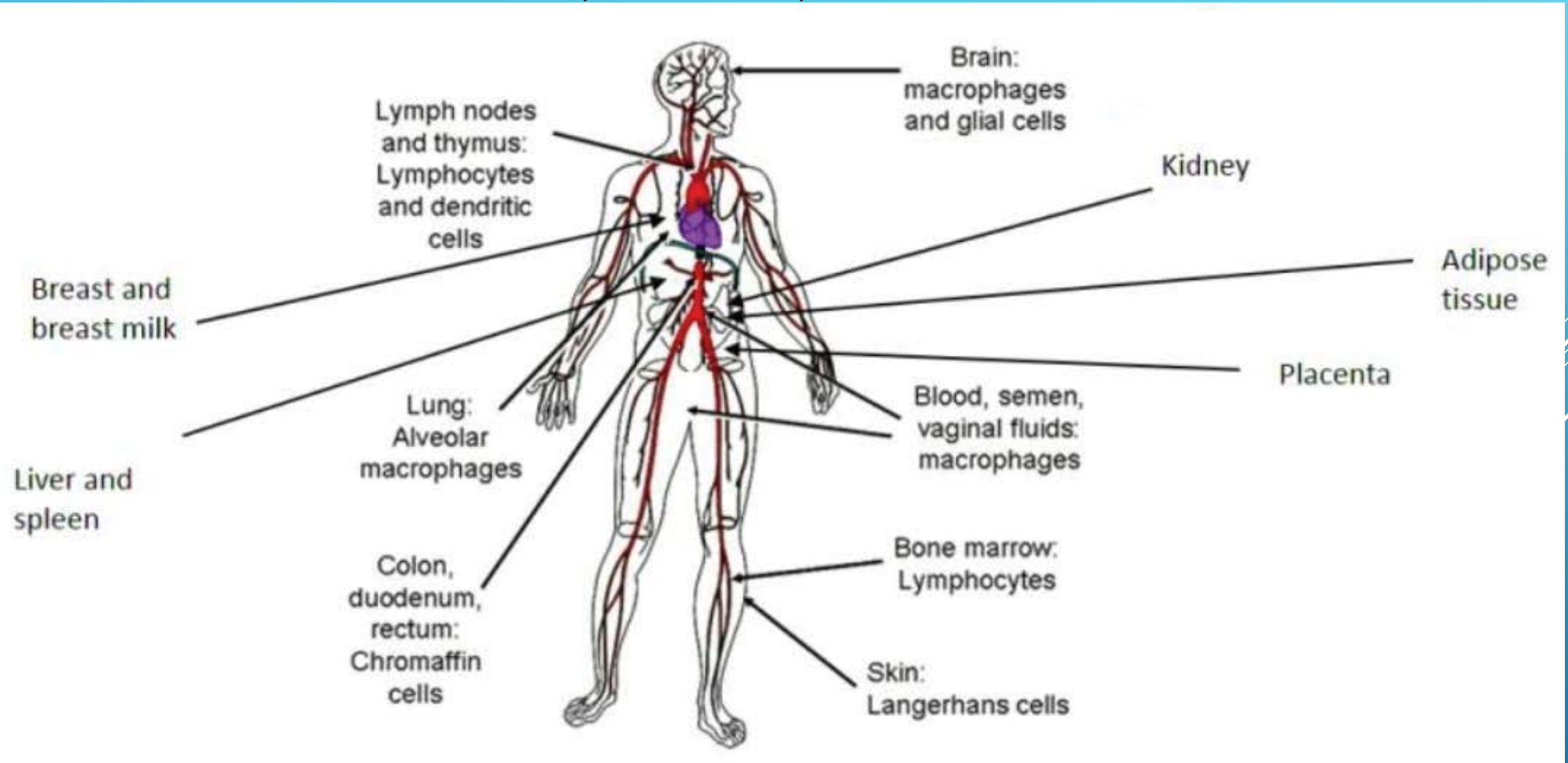
- Removal of ART results in rebound viremia within 3 weeks
- Occurs even in the setting of long-term viral suppression



Modified from Thompson C 2017

WHY DOES REBOUND OCCUR?

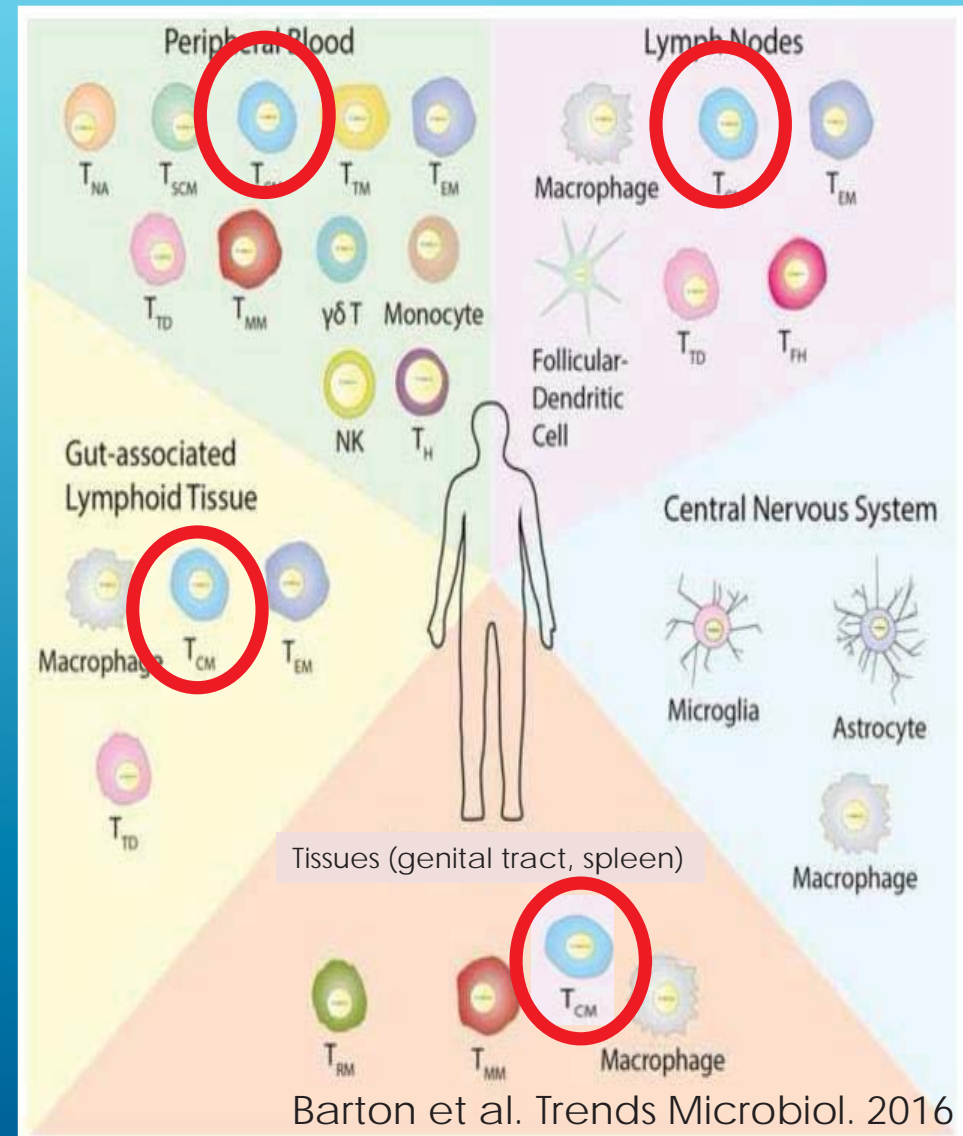
ANATOMICAL SITES OF HIV (DNA/RNA)



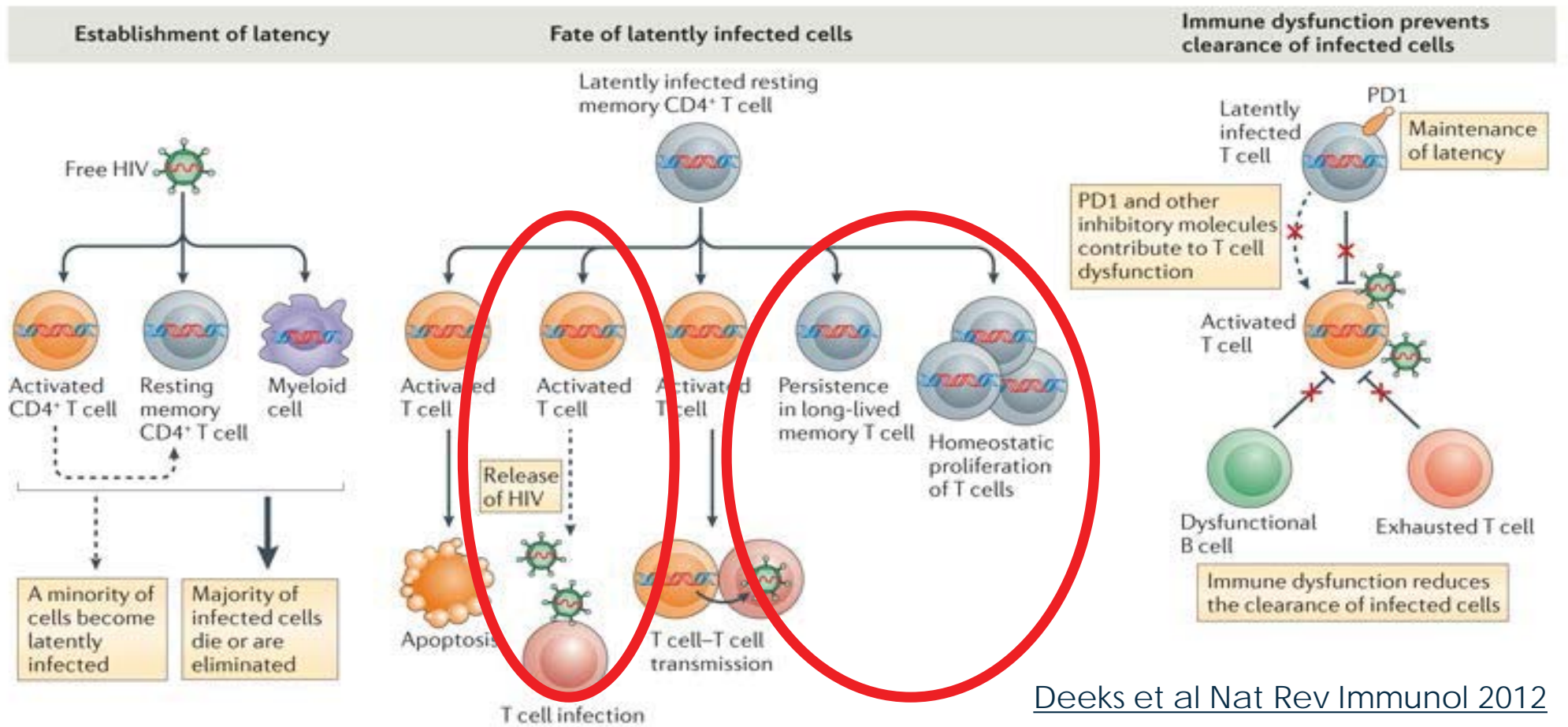
Adapted from Wong et al Curr Opin HIV AIDS 2016, Holdrych CROI 2018

THE VIRAL RESERVOIR: *A CENTRAL PROBLEM TO CURE*

- Sequestered anatomic sites
 - Genital tract
 - Central nervous system
- Central memory T cells (T follicular helper cells); quiescent
- Lymphoid organs
 - Lymph nodes
 - Spleen
 - Gastrointestinal tract



MULTIPLE MECHANISMS OF HIV PERSISTENCE



Deeks et al Nat Rev Immunol 2012

LOW LEVEL REPLICATION OR CLONAL EXPANSION?

LOW LEVEL REPLICATION (ARVs)

- Evidence of viral evolution in anatomic compartments
- ARV intensification with INSTI results in increased 2-LTR circles
- Imaging evidence of active HIV replication in LN under suppressive ARV therapy
- Lower drug concentrations in anatomical sites?

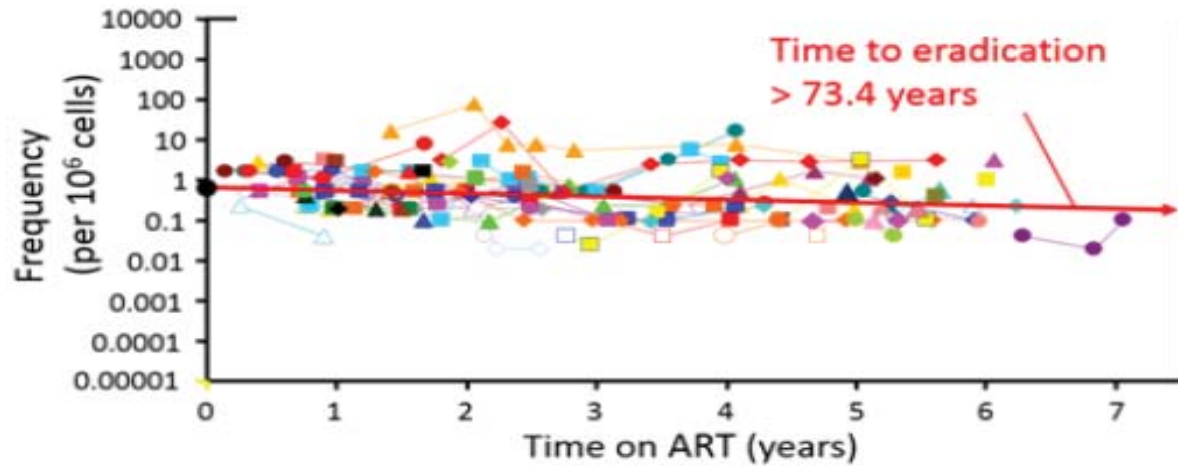
20+ references

CLONAL EXPANSION (IMSSs)

- No evidence of viral evolution in anatomic compartments
- ARV intensification does not change the size of the viral reservoir
- Evidence of clonal expansion in lymph node
- Absence of breakthrough resistance at a population level
- Drug concentration targets unknown at a cellular level

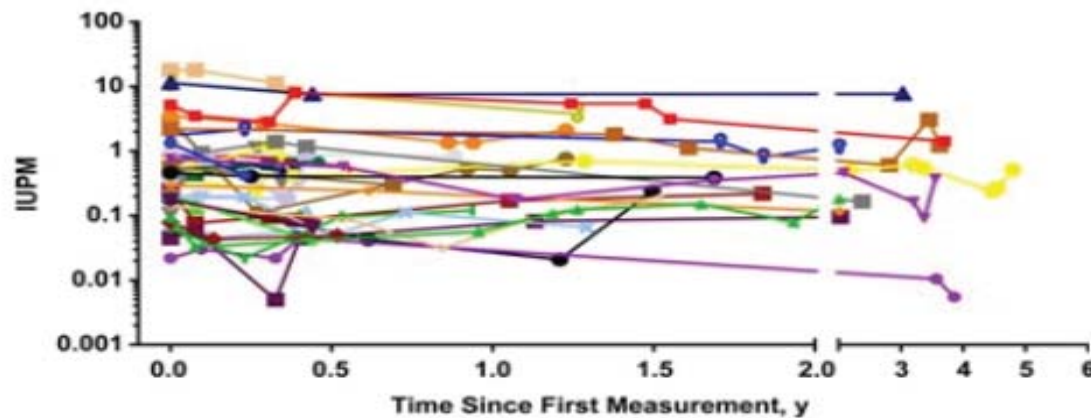
20+ references

WHATEVER IS RESPONSIBLE, THE VIRAL RESERVOIR DECAYS SLOWLY



$t_{1/2} = 44$ months

Siliciano et al.,
Nature Med., 2003



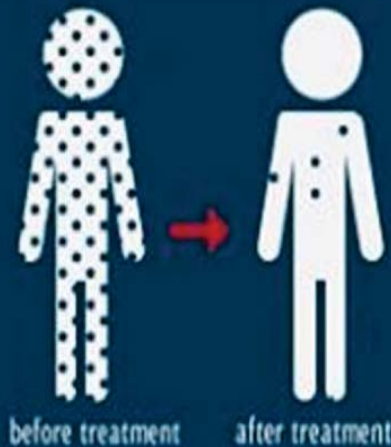
$t_{1/2} = 43$ months

Crook et al, JID 2015

CAN'T USE ARV TX ALONE FOR CURE: STRATEGIES FOR A CURE

FUNCTIONAL CURE

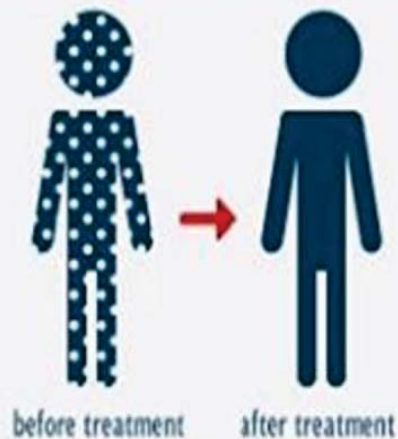
When the level of HIV particles in an infected person's body has been reduced to such an extremely low level that the person can stop treatment and not worry about the disease rebounding and damaging his immune system or body.



STERILIZING CURE

Eradication of HIV

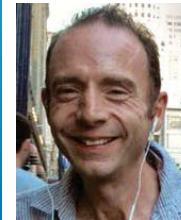
When every last particle of HIV has been destroyed or cleared out from an infected person's body.



Immune-mediated control of HIV infection (eg "Elite Controllers")

Withdraw ARV without rebound (eg remission after cancer therapy)

Increase inflammation/comorbidities?




N=1: Timothy Brown ("Berlin Patient")

SCT with CCR5 $\Delta 32\Delta 32$ + GVH Disease

Must remove virus AND target cells

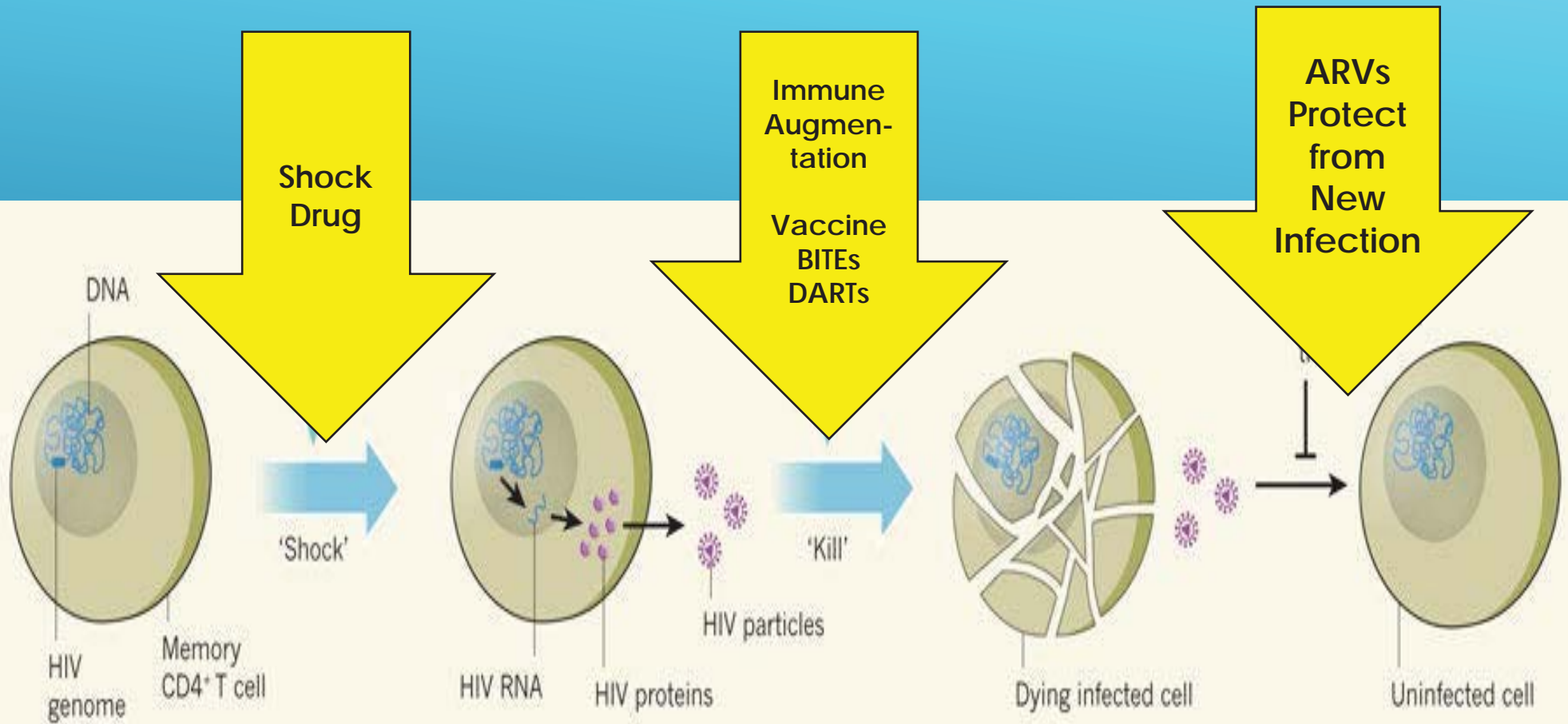
STRATEGIES FOR A CURE

Find and shrink the size of the HIV reservoir

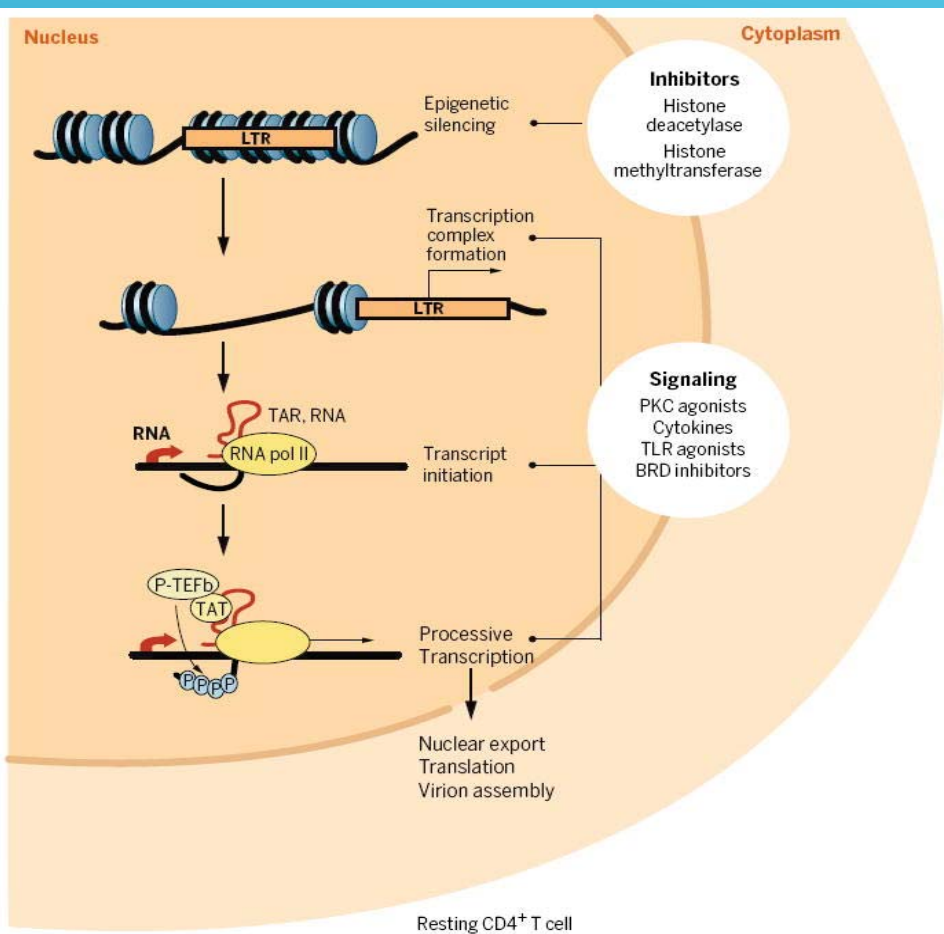
- ▶ Reduce seeding of the latent pool with early ARV therapy (eg Febig 1/2)
- ▶ Reverse latency (shock and kill) 
- ▶ Suppress latency (block and lock)
- ▶ Increase HIV-specific immune function (vaccines)
- ▶ Immune checkpoint blockade (antibodies – eg anti-PD1 Ab)
- ▶ Gene therapy targeting for the virus or host (CCR5 knockout/knockdown)
- ▶ Stem cell transplant

Combination Therapy Will Be Necessary

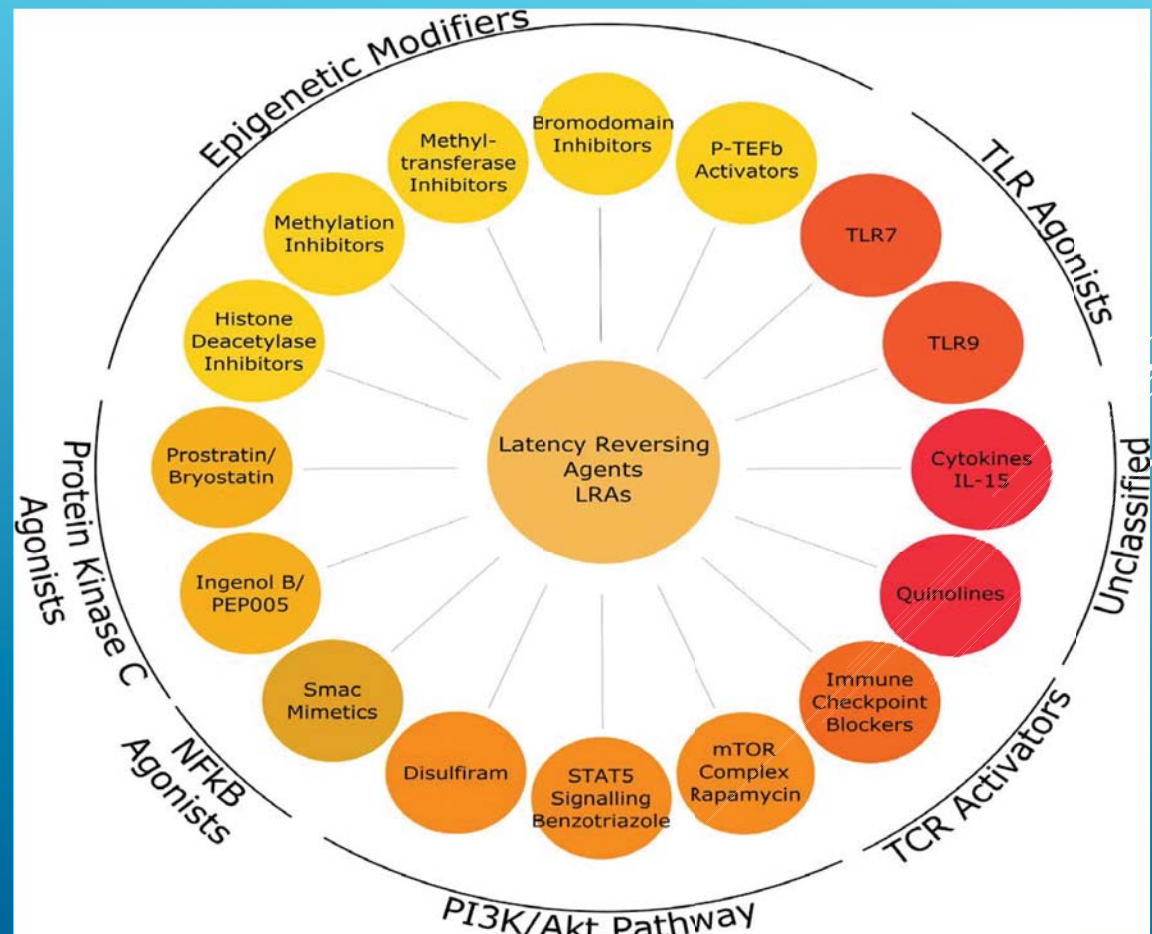
SHOCK AND KILL STRATEGY



SHOCK APPROACHES

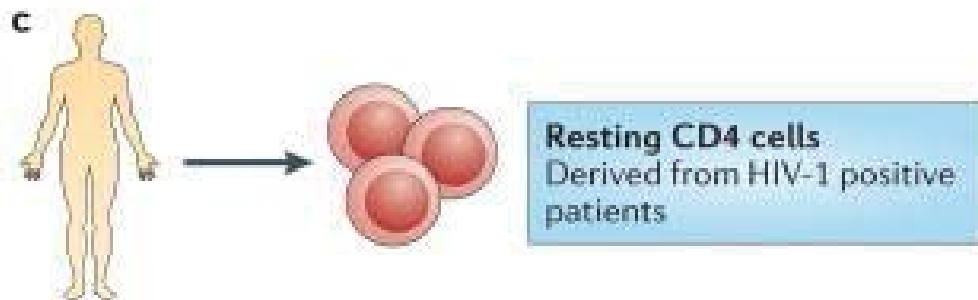
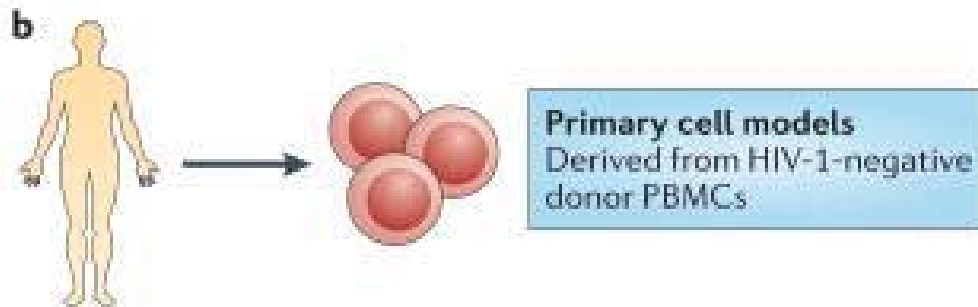


Margolis et al. Science. 2016



Kim et al. Cell Host and Microbe, 2018

CURRENT SYSTEMS TO STUDY HIV LATENCY



d Humanized SCID mouse



Irradiate SCID mouse,
transplanted with
human thymus and
foetal liver fragments

e Humanized BLT mouse



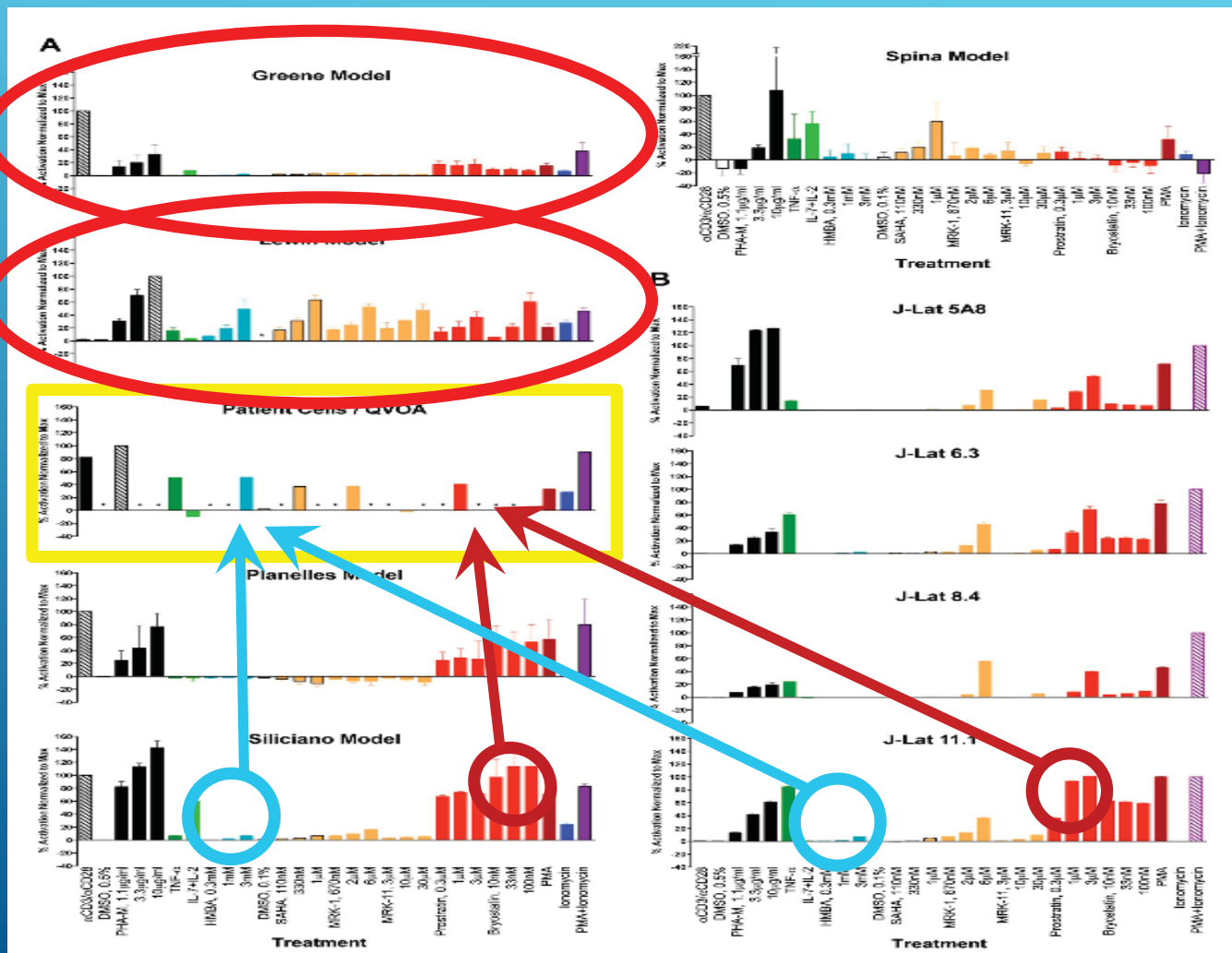
Irradiate NSG mice,
transplanted with foetal
human thymus and liver
tissue, and CD34⁺ stem
cells

f Macaque



Infect animals with:
SIV
RT-SHIV
SHIV

CHALLENGES WITH CELL MODELS



- 5 T cell models
- 4 J-Lat models
- 1 patient cells
- 13 stimuli
- no single model captured patient cell response
- PKC agonists and PHA reactivated HIV across models; drugs in most other classes did not

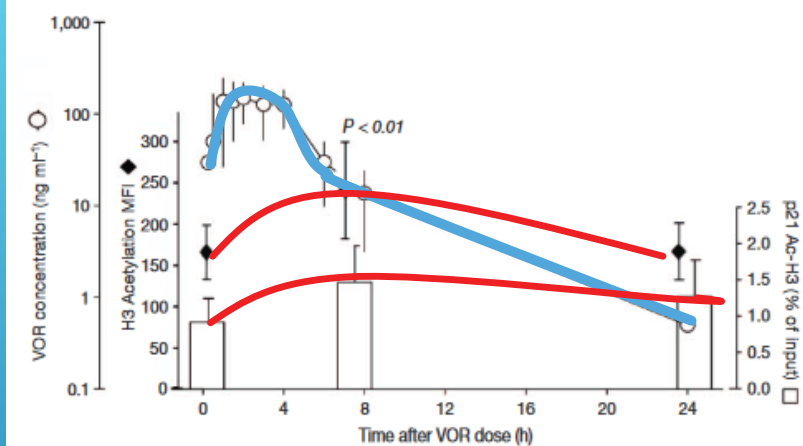
Spina et al. An in-depth comparison of latent HIV-1 reactivation in multiple cell model systems and resting CD4+ T cells from aviremic patients. PLoS Pathog. 2013;9(12):e1003834.

CHALLENGES WITH EXTRAPOLATION TO CLINICAL TRIALS

Nature 2012

Administration of vorinostat disrupts HIV-1 latency in patients on antiretroviral therapy

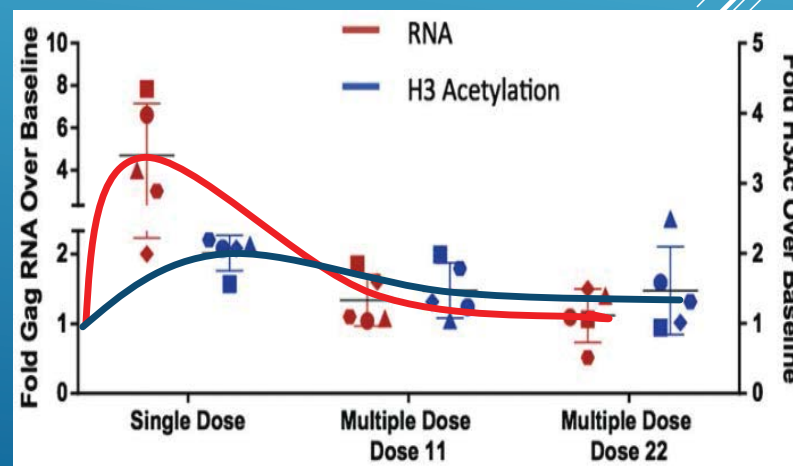
N. M. Archin¹, A. L. Liberty¹, A. D. Kashuba¹, S. K. Choudhary¹, J. D. Kuruc¹, A. M. Crooks¹, D. C. Parker¹, E. M. Anderson², M. F. Kearney², M. C. Strain³, D. D. Richman³, M. G. Hudgens¹, R. J. Bosch⁴, J. M. Coffin², J. J. Eron¹, D. J. Hazuda⁵ & D. M. Margolis¹



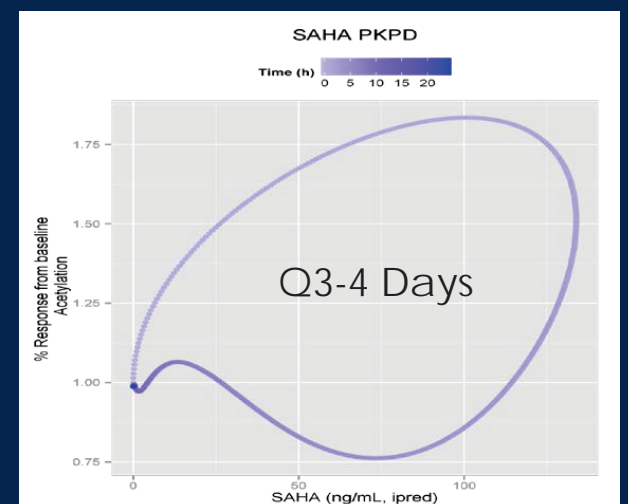
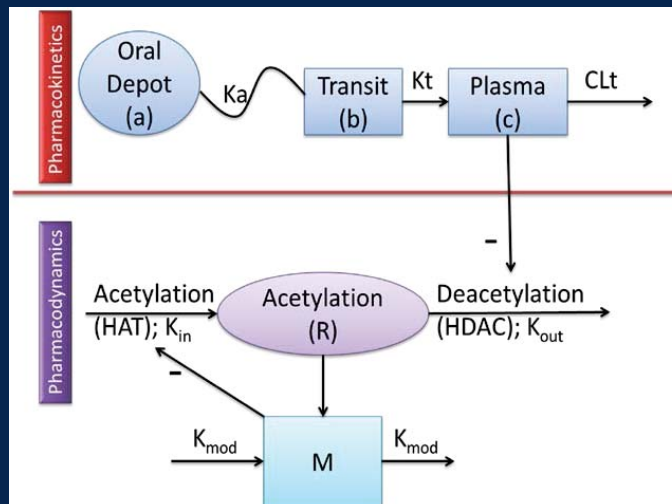
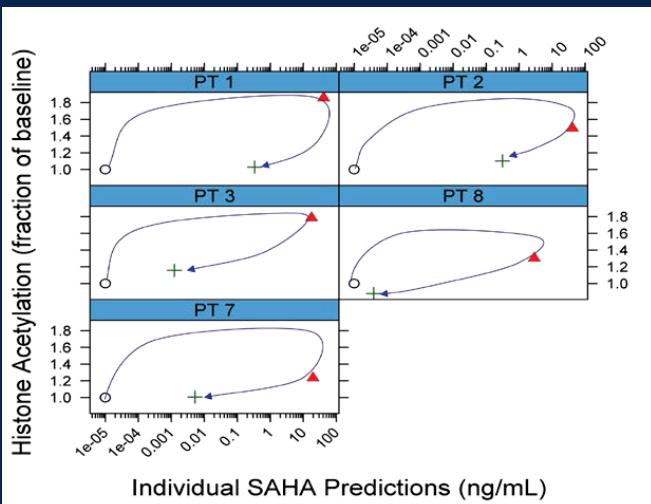
HIV-1 Expression Within Resting CD4⁺ T Cells After Multiple Doses of Vorinostat

JID 2014

Nancy M. Archin,¹ Rosalie Bateson,¹ Manoj K. Tripathy,¹ Amanda M. Crooks,¹ Kuo-Hsiung Yang,¹ Noelle P. Dahl,¹ Mary F. Kearney,² Elizabeth M. Anderson,² John M. Coffin,^{2,3} Matthew C. Strain,⁴ Douglas D. Richman,⁴ Kevin R. Robertson,¹ Angela D. Kashuba,¹ Ronald J. Bosch,⁵ Daria J. Hazuda,⁶ Joann D. Kuruc,¹ Joseph J. Eron,¹ and David M. Margolis¹



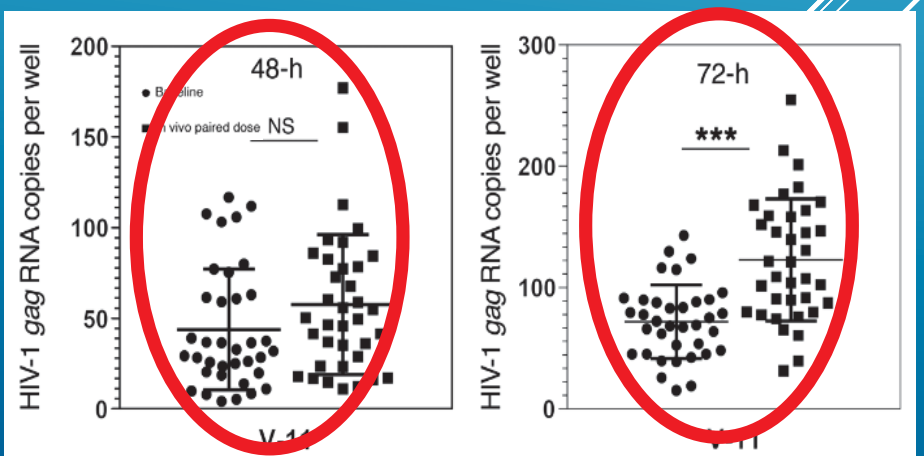
USING PHARMACOMETRICS TO OPTIMIZE DOSING



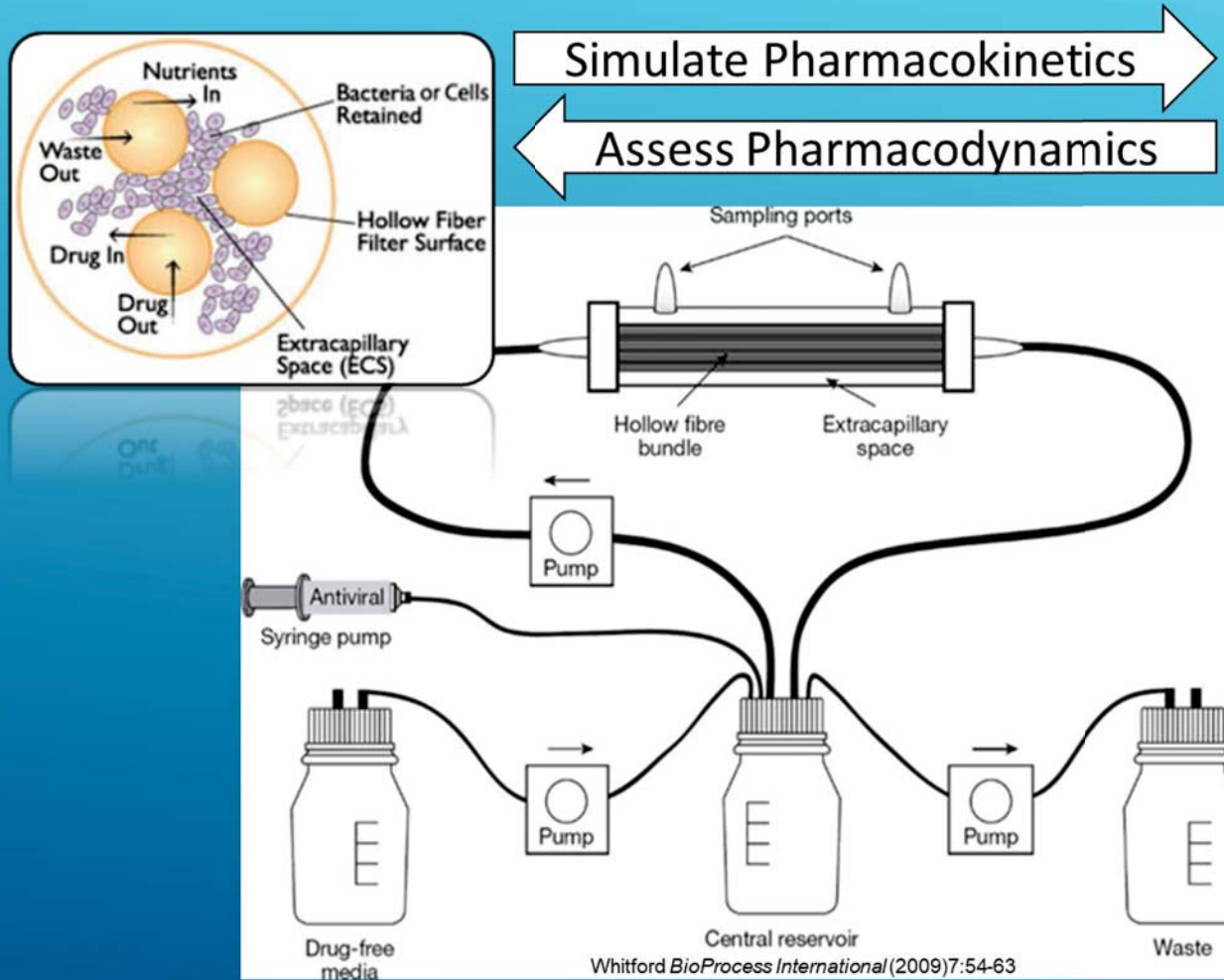
Interval dosing with the HDAC inhibitor vorinostat effectively reverses HIV latency

JCI 2017

Nancie M. Archin,^{1,2} Jennifer L. Kirchherr,¹ Julia A.M. Sung,^{1,2} Genevieve Clutton,^{1,3} Katherine Sholtis,¹ Yinyan Xu,¹ Brigitte Allard,¹ Erin Stuelke,¹ Angela D. Kashuba,⁴ Joann D. Kuruc,^{1,2} Joseph Eron,^{1,2,5} Cynthia L. Gay,^{1,2} Nilu Goonetilleke,^{1,2} and David M. Margolis^{1,2,5,6}



IMPROVING STATIC IN VITRO SYSTEM PREDICTABILITY



- Central reservoir mimics *in vivo* **circulation**
- Syringe pump infuses drug treated media to mimic *in vivo* drug **absorption**
- Media pumps dilute drug treated media in central reservoir to mimic *in vivo* drug **clearance**

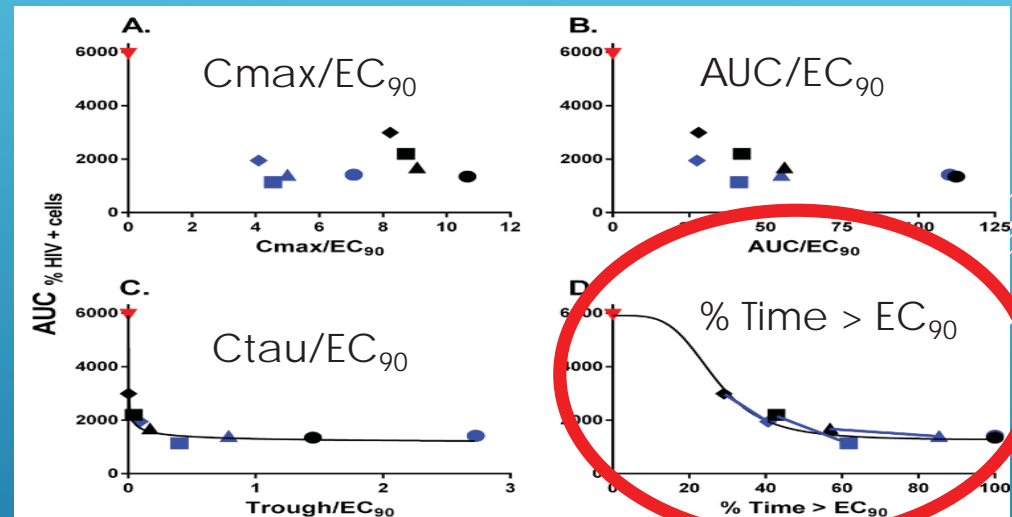
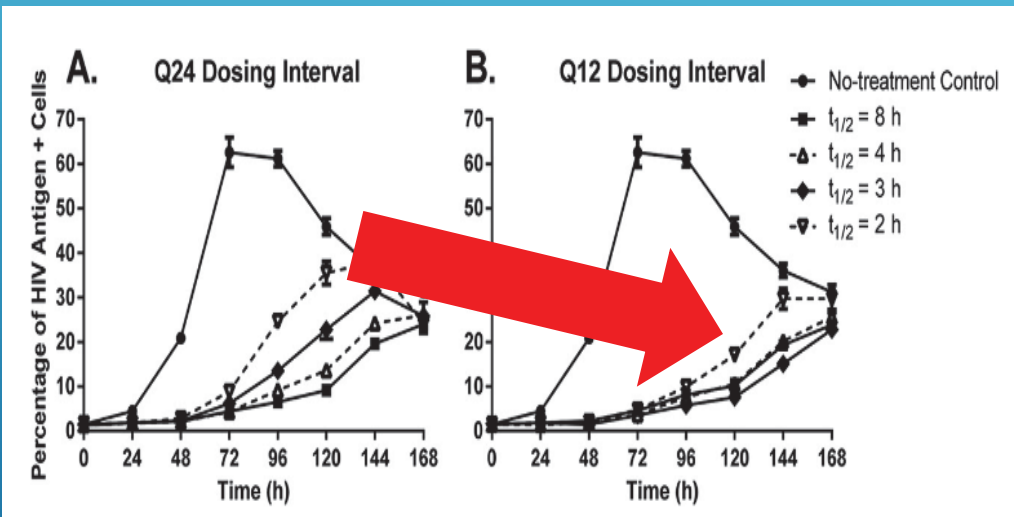
Figure: Graphical Representation of a Hollow Fiber Model (HFM). Whitford *BioProcess International* (2009)7:54-63

HOLLOW FIBER BIOREACTOR POTENTIAL

Pharmacokinetic Determinants of Virological Response to Raltegravir in the *In Vitro* Pharmacodynamic Hollow-Fiber Infection Model System

Ashley N. Brown, Jonathan R. Adams, Dodge L. Baluya, George L. Drusano

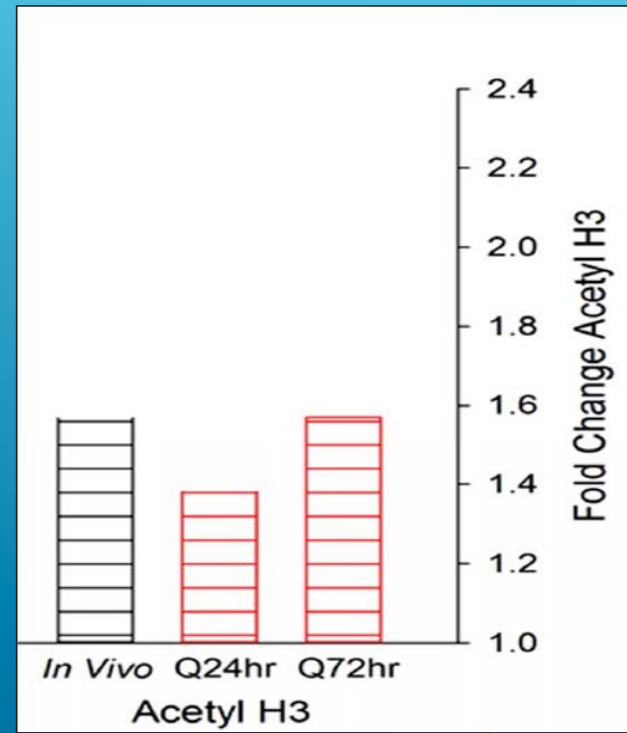
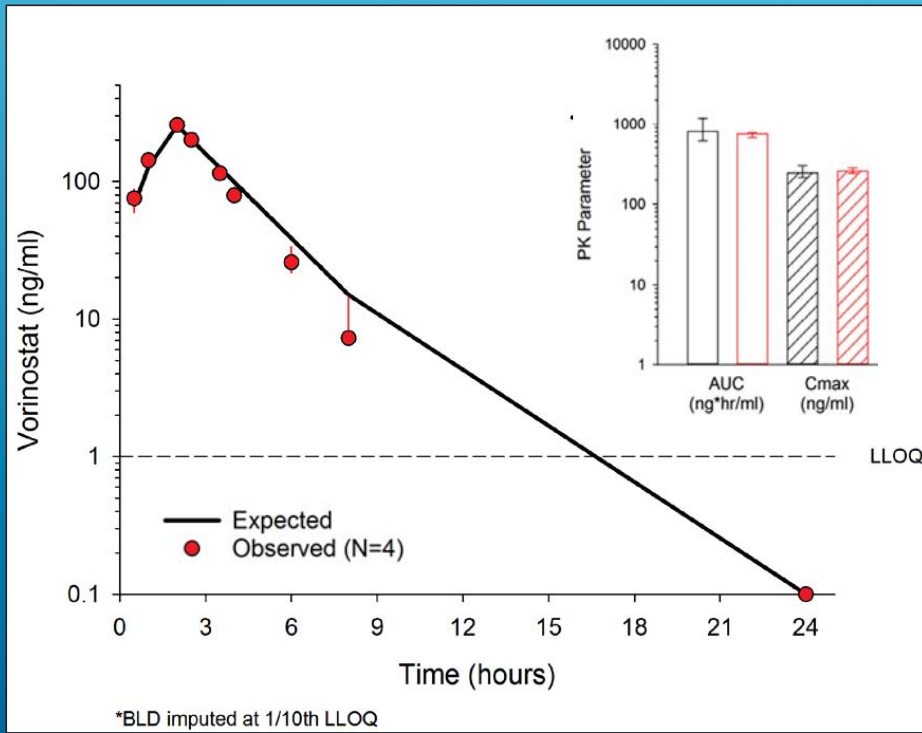
AAC 2015



PK/PD Predictions

- ▶ Meropenem for Pseudomonas
- ▶ Aztreonam+avibactam for MDR Enterobacteriaceae
- ▶ Oseltamivir for Influenza A
- ▶ Amprenavir+ritonavir for HIV

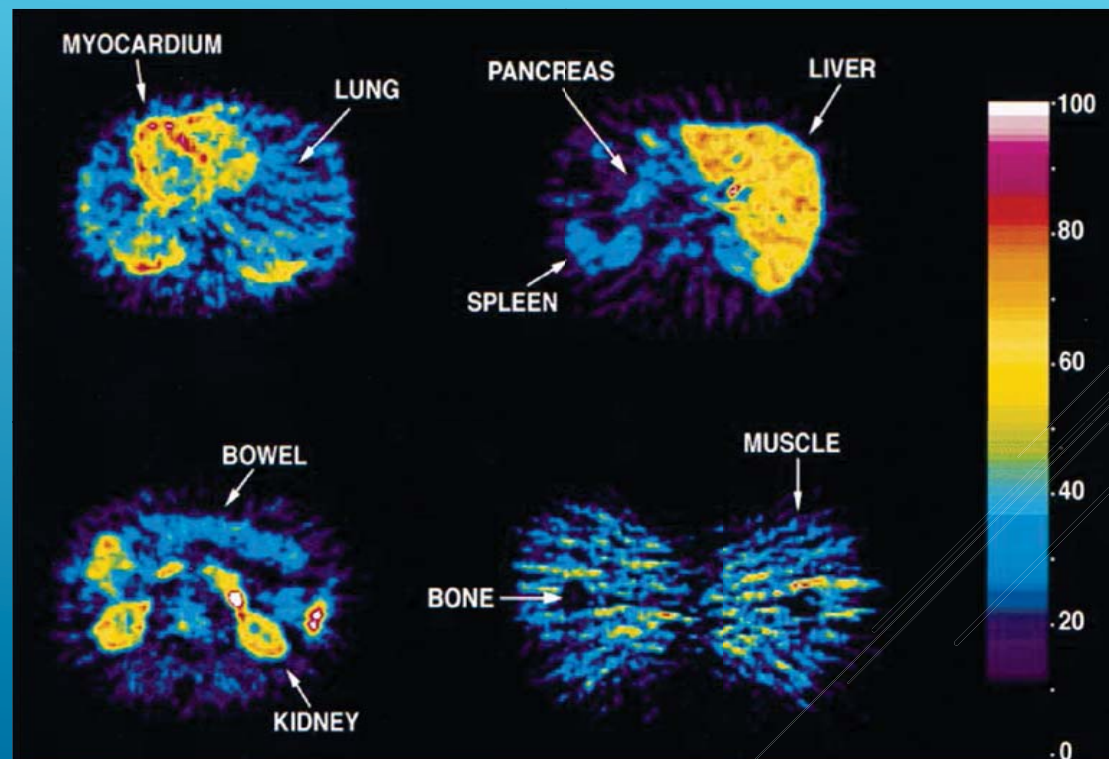
HOLLOW FIBER BIOREACTOR POTENTIAL



Archin JID 2014
Archin JCI 2017
Cottrell, unpublished 2018

CHALLENGES WITH TISSUE DISTRIBUTION/PHARMACOKINETICS: *LATENCY REVERSING AGENTS AND ANTIRETROVIRALS*

- ▶ Tissue distribution is heterogeneous
 - ▶ Tissue distribution is nonhomogeneous and tissue specific, with high inter-tissue and inter-subject variability
 - ▶ target site concentrations may substantially differ from plasma concentrations



Representative PET images of human subjects following the ¹⁸F-trovafoxacin. Muller AAC 2004

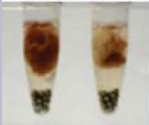
MEASURING TISSUE PHARMACOKINETICS FOR LRA AND ARV PK/PD

LC-MS methods of quantification

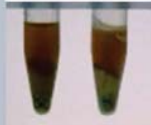
- Extraction of analyte from tissue homogenate
- Useful for providing initial information of averaged concentration
- Lack of spatial resolution
- intracellular+extracellular

Tissue Biopsies Homogenized

Before

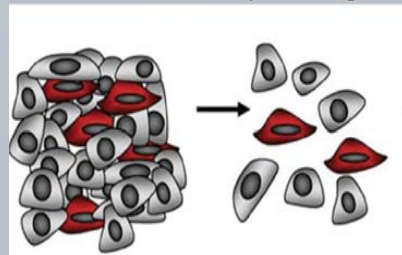


After



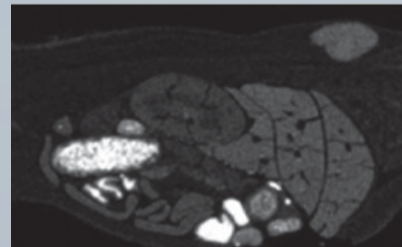
Enzymatic Digestion & Cell Isolation

- Concentrations decline with sample processing
- Lack of spatial resolution



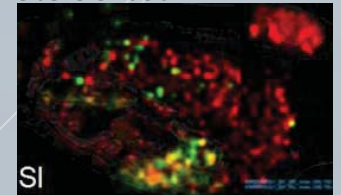
Traditional imaging techniques (QWBA, PET)

- Require radiolabels
- May not distinguish between parent and metabolite
- Challenging to evaluate multi-drug therapies
- Cellular resolution may not be possible



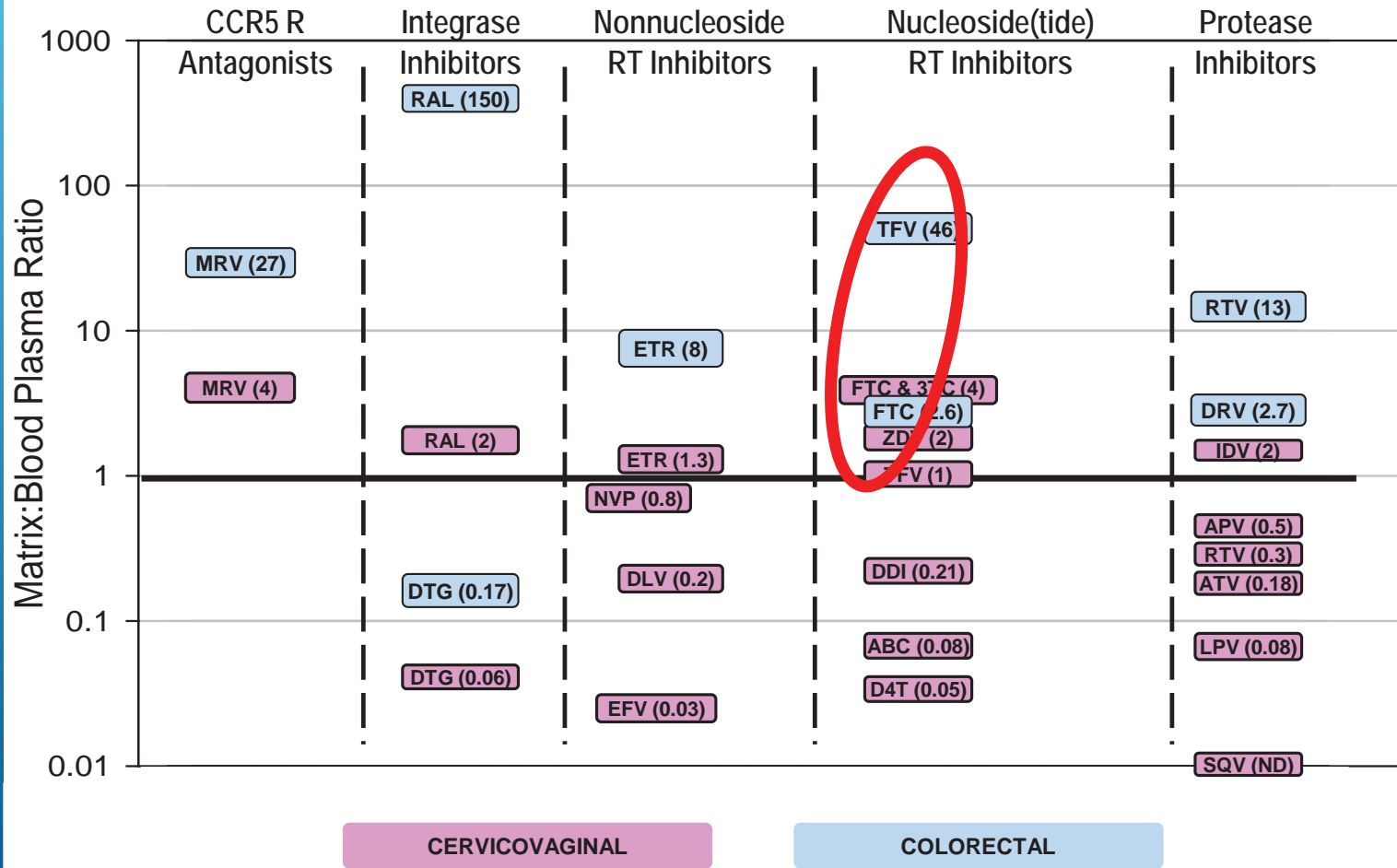
Mass Spectroscopy Imaging

- Only allows ex-vivo measures
- Spatial distribution of compounds
- Distinguishes between parent and metabolite
- Provide additional information on endogenous compounds and metabolites



UTILITY OF MUCOSAL TISSUE HOMOGENATES IN PREP

Cottrell et al 2015



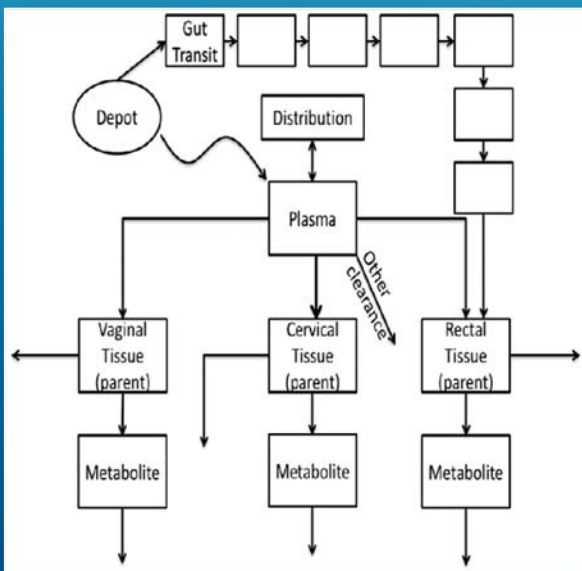
UTILITY OF MUCOSAL TISSUE HOMOGENATES IN PREP

A Translational Pharmacology Approach to Predicting Outcomes of Preexposure Prophylaxis Against HIV in Men and Women Using Tenofovir Disoproxil Fumarate With or Without Emtricitabine

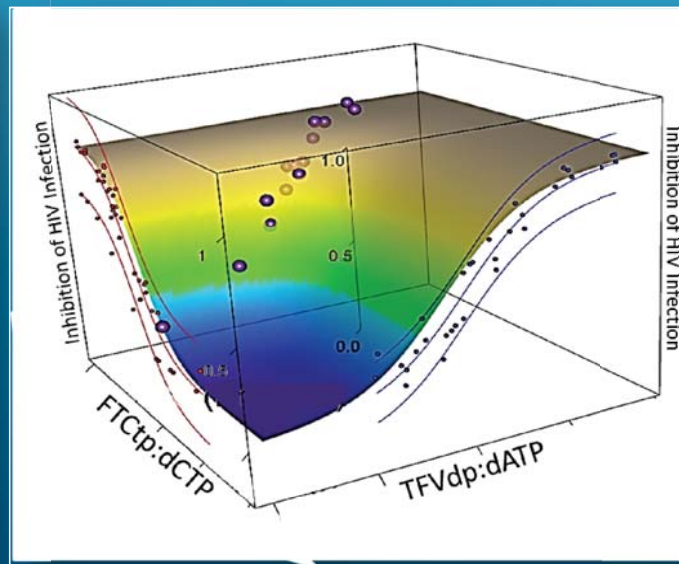
JID 2016

Mackenzie L. Cottrell,¹ Kuo H. Yang,² Heather M. A. Prince,³ Craig Sykes,¹ Nicole White,¹ Stephanie Malone,¹ Evan S. Dellon,³ Ryan D. Madanick,³ Nicholas J. Shaheen,³ Michael G. Hudgens,⁴ Jacob Wulff,⁴ Kristine B. Patterson,³ Julie A. E. Nelson,⁵ and Angela D. M. Kashuba¹

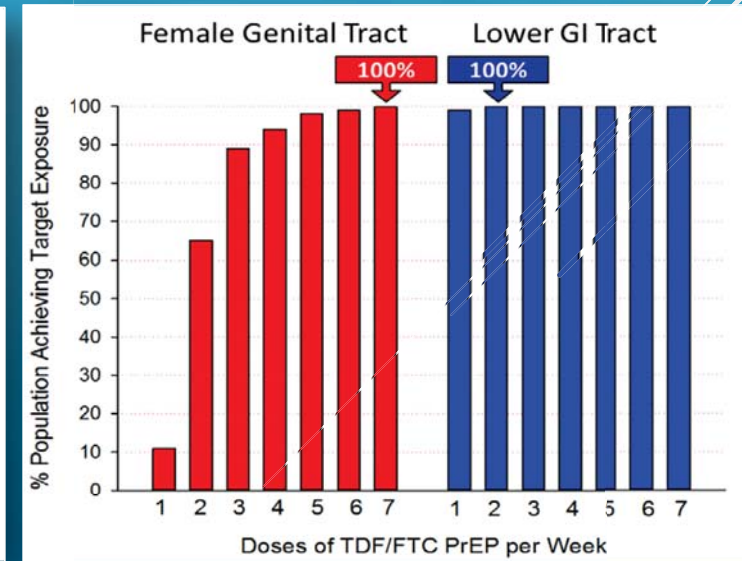
PK Model of TFVdp and FTCtp in Mucosal Tissues



In Vitro Cellular Targets for Efficacy/Synergy

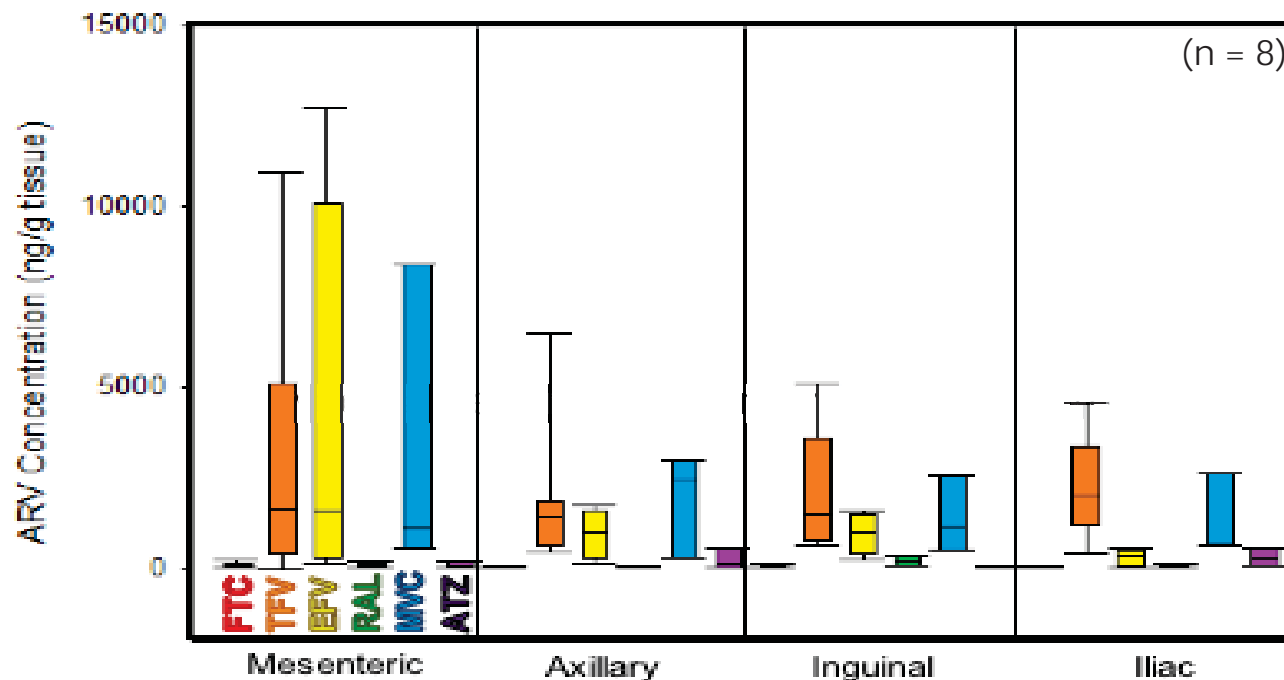


Dose Frequency for HIV Protection



UTILITY OF TISSUE HOMOGENATES LIMITED IN CURE LN: WHAT CONCENTRATION IS NEEDED FOR EFFICACY?

LC-MS/MS ARV Concentration Analysis in Lymph Nodes



Need a better understanding of tissue pk/pd



Emtricitabine (FTC) Raltegravir (RAL)
Tenofovir (TFV) Maraviroc (MVC)
Efavirenz (EFV) Atazanavir (ATZ)

Burgunder et al. Unpublished data. 2018

ENZYMATIC CELL DIGESTION

Defining total-body AIDS-virus burden with implications for curative strategies

Nature Med 2017

Jacob D Estes¹, Cissy Kityo², Francis Ssali², Louise Swainson³, Krystelle Nganou Makamdop⁴, Gregory Q Del Prete¹, Steven G Deeks⁵, Paul A Luciw⁶, Jeffrey G Chipman⁷, Gregory J Beilman⁷ , Torfi Hoskuldsson⁷, Alexander Khoruts⁸, Jodi Anderson⁸, Claire Deleage¹, Jacob Jasurda⁸, Thomas E Schmidt⁸, Michael Hafertepe⁸, Samuel P Callisto⁸ , Hope Pearson⁸, Thomas Reimann⁸, Jared Schuster⁸, Jordan Schoepfoerster⁸, Peter Southern⁹, Katherine Perkey⁹, Liang Shang⁹, Stephen W Wietgreffe⁹, Courtney V Fletcher¹⁰, Jeffrey D Lifson¹, Daniel C Douek⁴, Joseph M McCune³, Ashley T Haase⁹ & Timothy W Schacker⁸

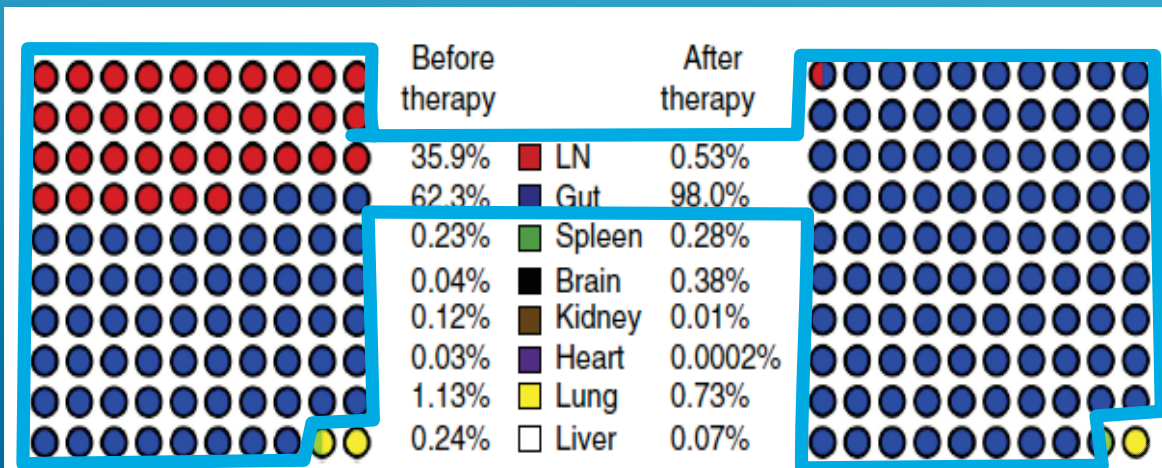


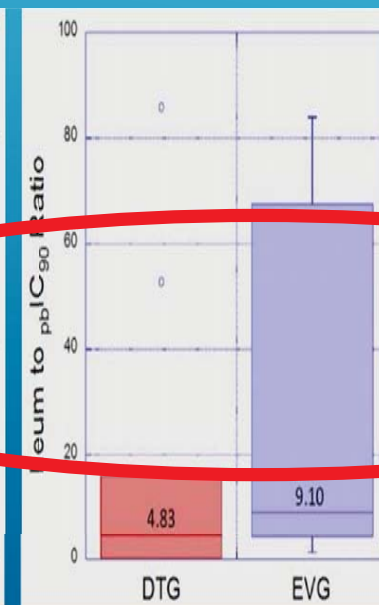
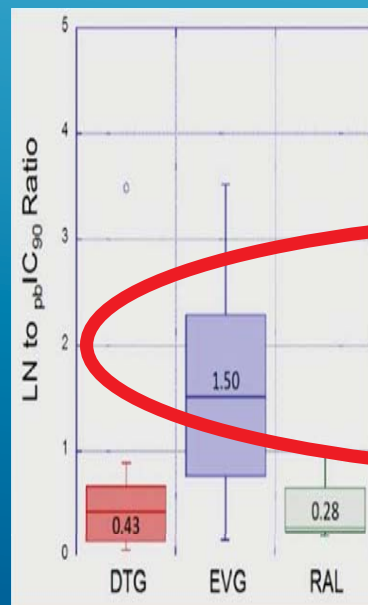
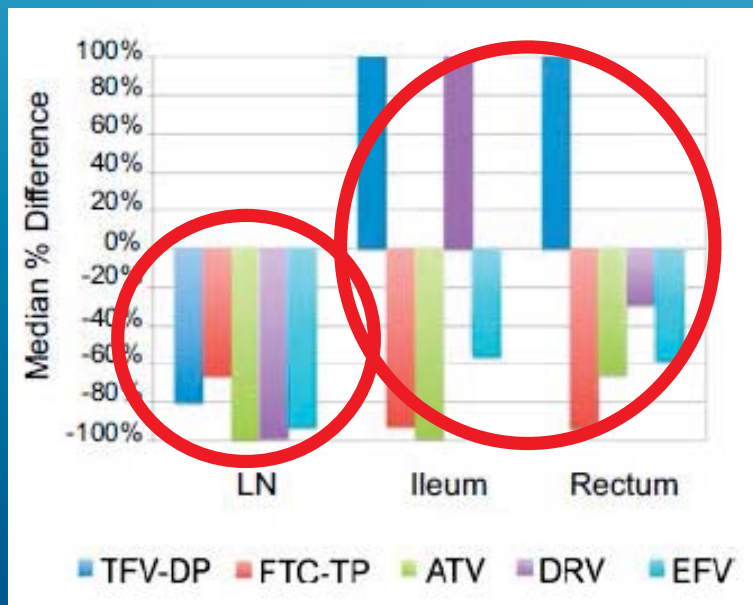
Figure 1 Graphical representation of the proportion of vRNA⁺ cells in each organ system before and during suppressive ART.

ENZYMATIC CELL DIGESTION

PNAS 2014

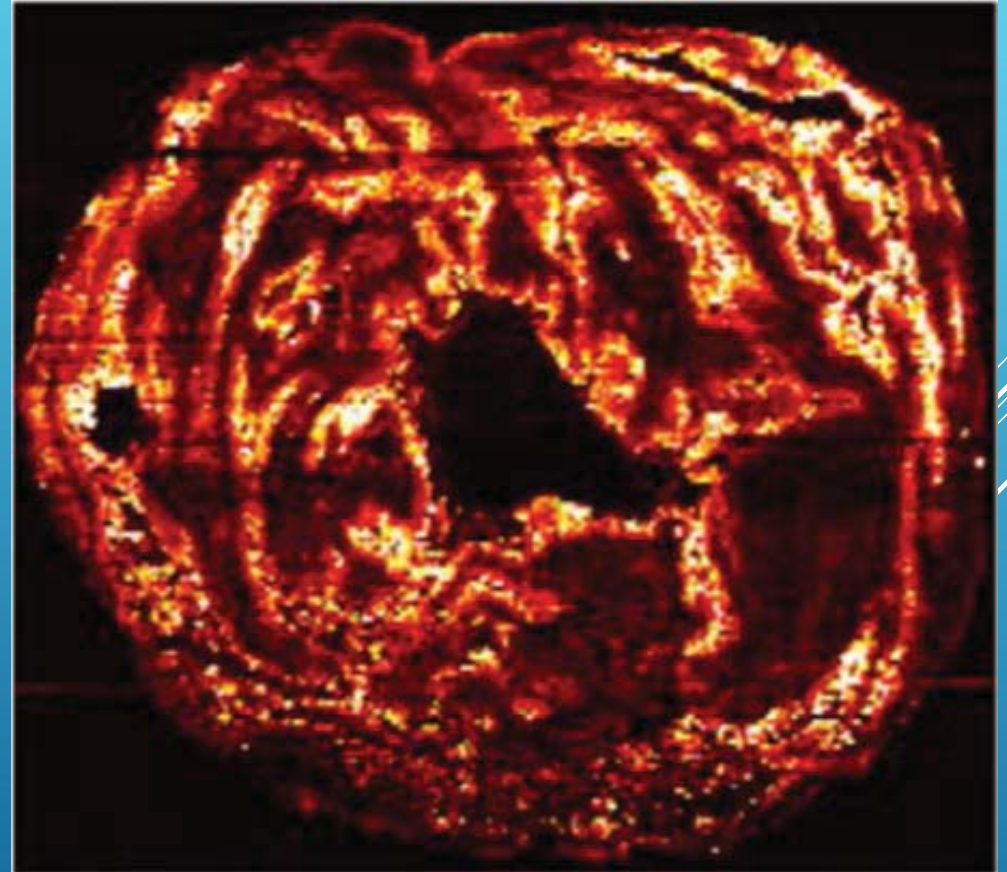
Persistent HIV-1 replication is associated with lower antiretroviral drug concentrations in lymphatic tissues

Courtney V. Fletcher^a, Kathryn Staskus^{b,1}, Stephen W. Wietgreffe^b, Meghan Rothenberger^c, Cavan Reilly^d, Jeffrey G. Chipman^e, Greg J. Beilman^e, Alexander Khoruts^c, Ann Thorkelson^c, Thomas E. Schmidt^c, Jodi Anderson^c, Katherine Perkey^b, Mario Stevenson^f, Alan S. Perelson^g, Daniel C. Douek^h, Ashley T. Haase^b, and Timothy W. Schacker^{c,2}



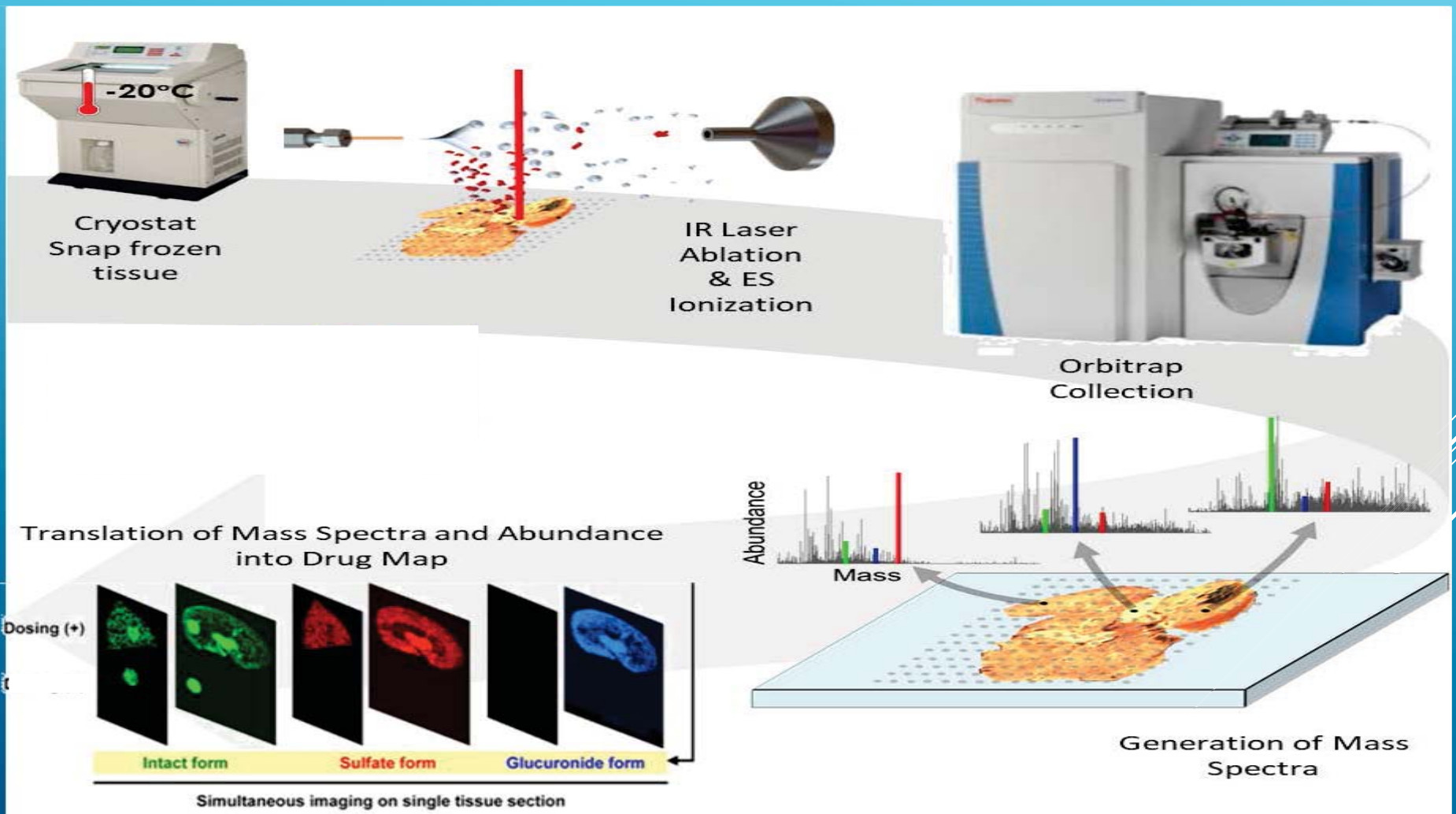
MASS SPECTROSCOPY IMAGING

- Could HIV persistence in reservoirs be due to inadequate ARV distribution?
- Will anti-latency therapies reach all tissue sites ?



Thompson, et al. AAC. 2015, 59(5)

QUANTITATIVE IR MALDESI

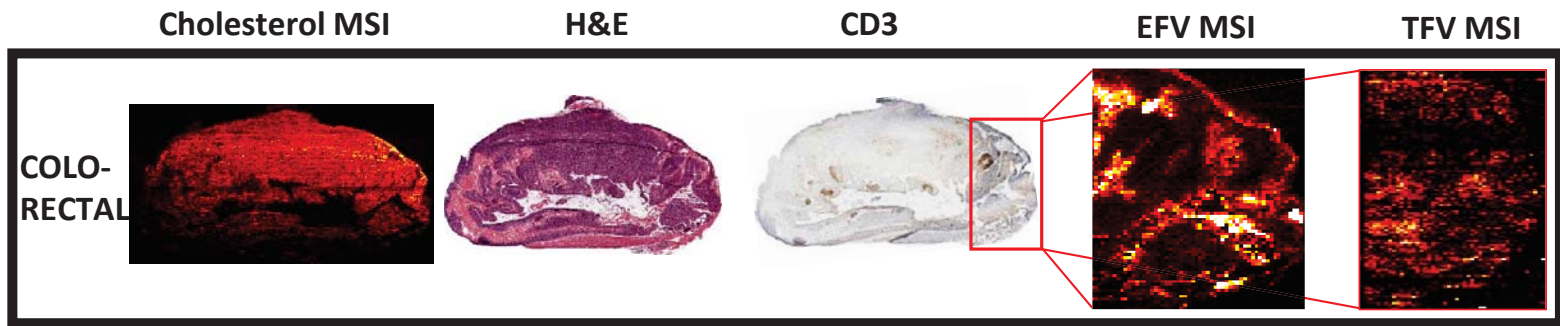


MSI OF ARVS IN TISSUES: *DRUG-SPECIFIC DISTRIBUTION*

↑ Conc



↓ Conc

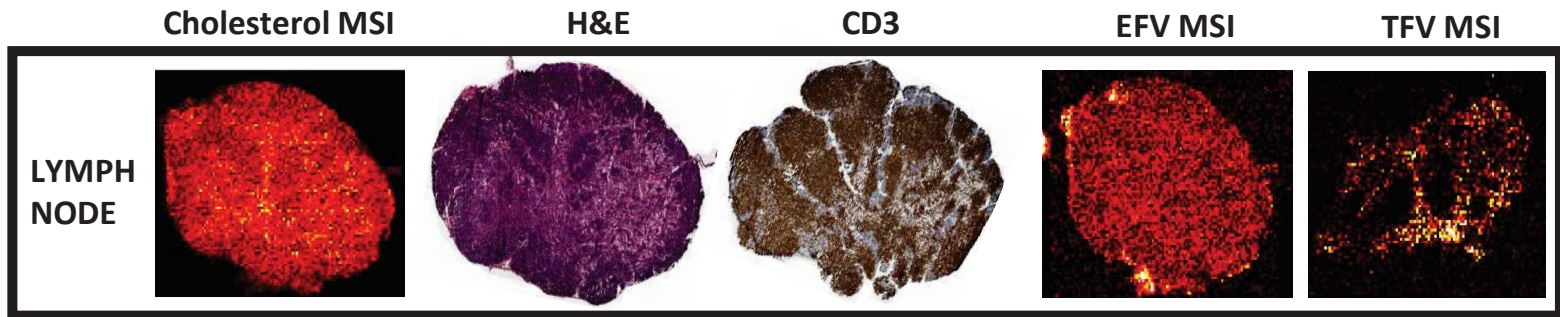


localized to mucosa, lamina propria

Intra-tissue concentration gradient

10 fold

3 fold



localized to capsule, few follicles

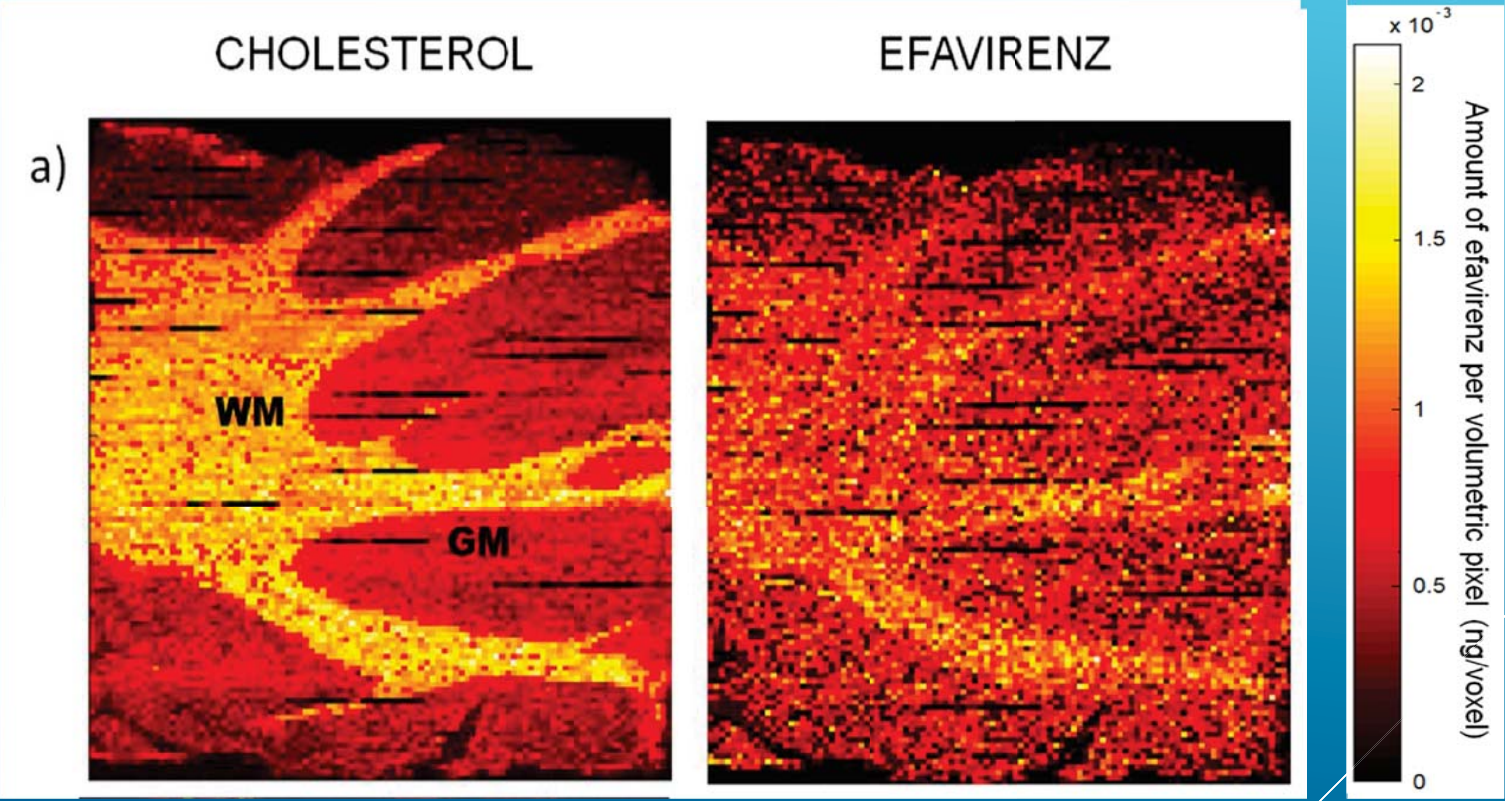
medullary sinuses

Intra-tissue concentration gradient

3 fold

17 fold

ANATOMIC RESERVOIR MSI: *WHITE VERSUS GREY MATTER*



PANOBINOSTAT (PANO- HDAC INHIBITOR):

IN VITRO EFFICACY – IN VIVO FAILURE

*Is the drug where it needs to be?
IR MALDESI Visualizes PANO In Vascular Spaces (70%) and
Follicles (30%)*

cells

AAC
Journals.AGM.org

Ex Vivo Bioactivity and HIV-1 Latency Reversal by Ingenol Dibenzoate and Panobinostat in Resting CD4⁺ T Cells from Aviremic Patients

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humanized mice **Retrovirology**

Tsai et al. *Retrovirology* (2016) 13:36
DOI 10.1186/s12977-016-0268-7

In vivo analysis of the effect of panobinostat on cell-associated HIV RNA and DNA levels and latent HIV infection

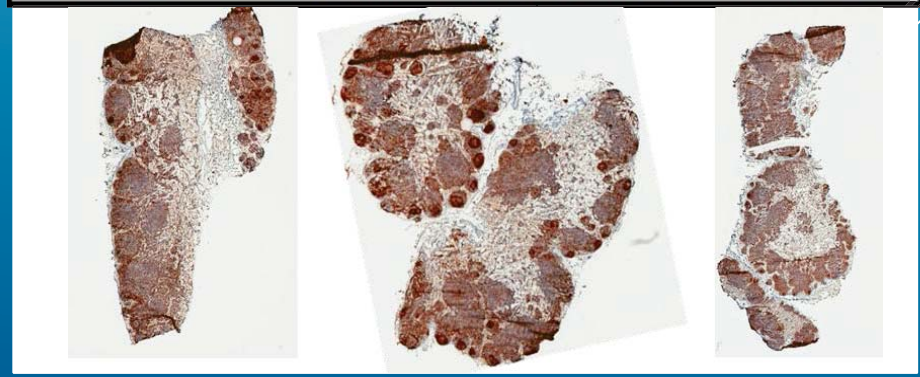
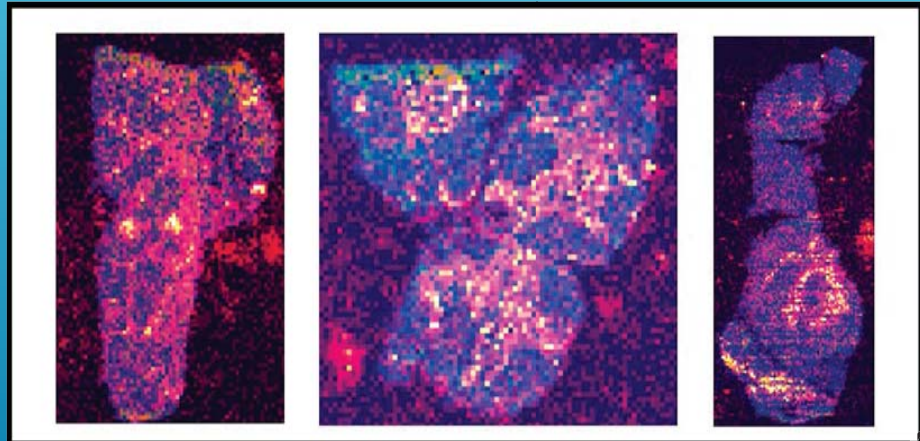
Perry Tsai¹, Guoxin Wu², Caroline E. Baker¹, William O. Thayer¹, Rae Ann Spagnuolo¹, Rosa Sanchez², Stephanie Barrett², Bonnie Howell², David Margolis¹, Daria J. Hazuda², Nancie M. Archin^{1*} and J. Victor Garcia^{1*}

HIV+ Patients

Panobinostat, a histone deacetylase inhibitor, for latent-virus reactivation in HIV-infected patients on suppressive antiretroviral therapy: a phase 1/2, single group, clinical trial

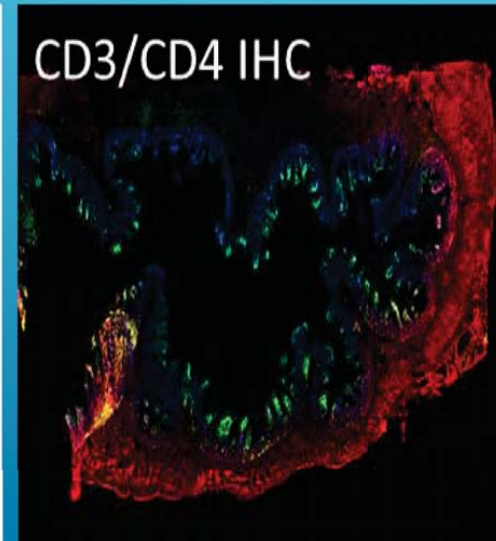
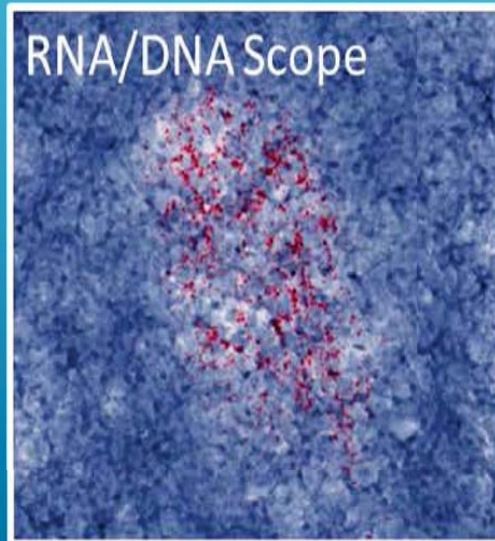
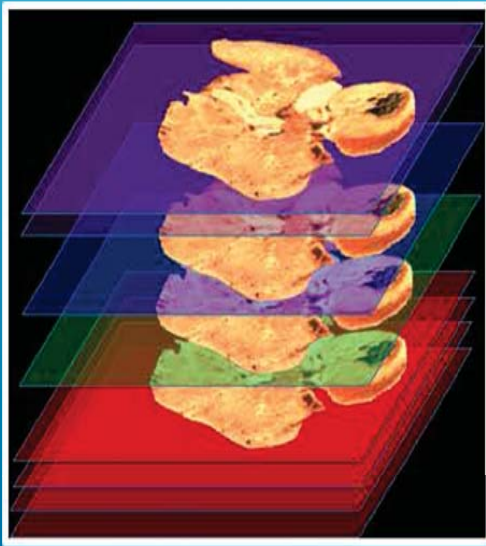
Thomas A Rasmussen, Martin Tolstrup, Christel R Brinkmann, Rikke Olesen, Christian Erikstrup, Ajantha Solomon, Anni Winckelmann, Sarah Palmer, Charles Dinarello, Maria Buzon, Mathias Lichterfeld, Sharon R Lewin, Lars Ostergaard, Ole S Sogaard

Summary
Background Activating the expression of latent virus is an approach that might form part of an HIV cure. We assessed *Lancet HIV* 2014; 1: e13-21



**IHC PD1
Staining of
NHP Lymph
Nodes**

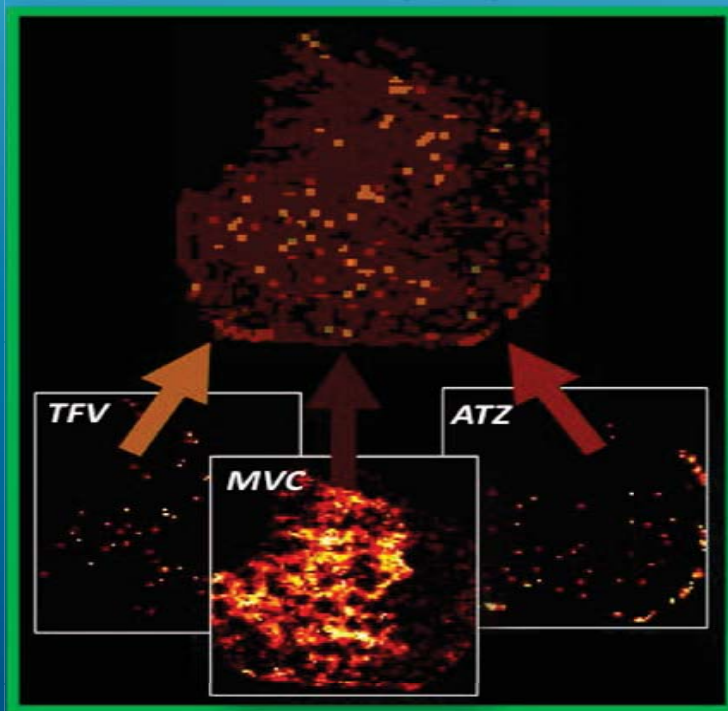
ADDITIONAL INSIGHTS WITH MSI



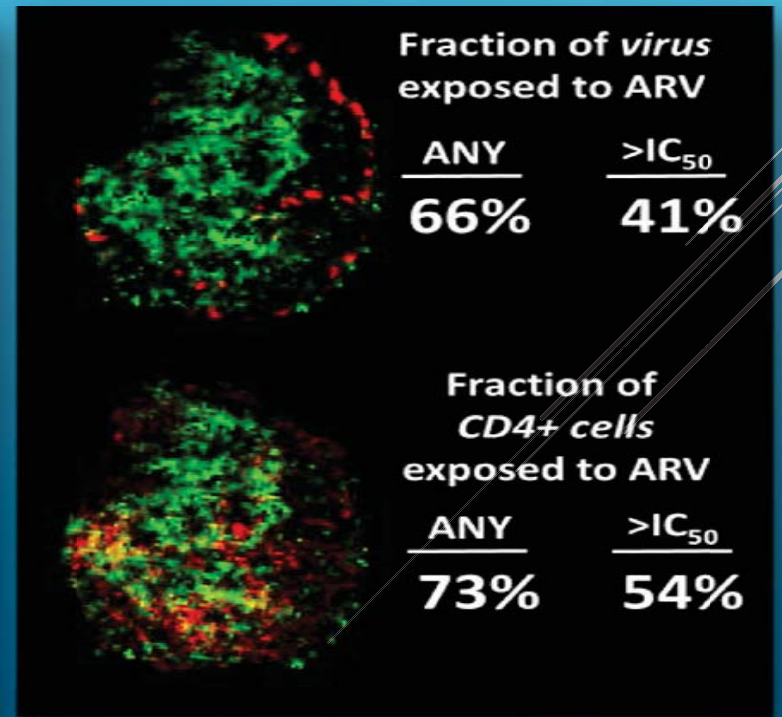
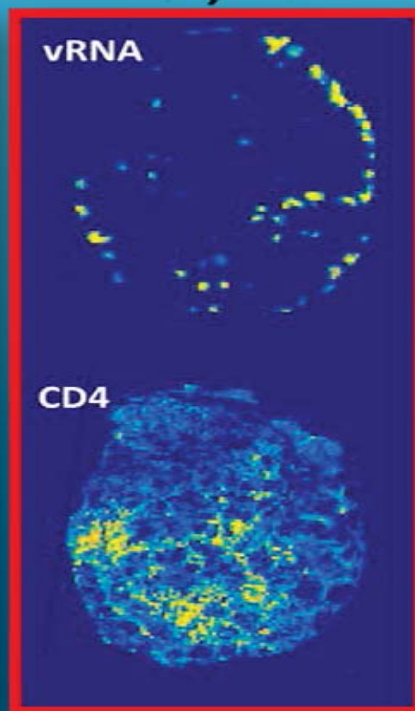
LYMPHOID TISSUE MSI: *LYMPH NODE OVERLAY FOR PK/PD?*

Total ARV Exposure in Lymph Node + Viral Expression or Target Cell Distribution = Image Fusion of ARV and vRNA or CD4+ T Cells

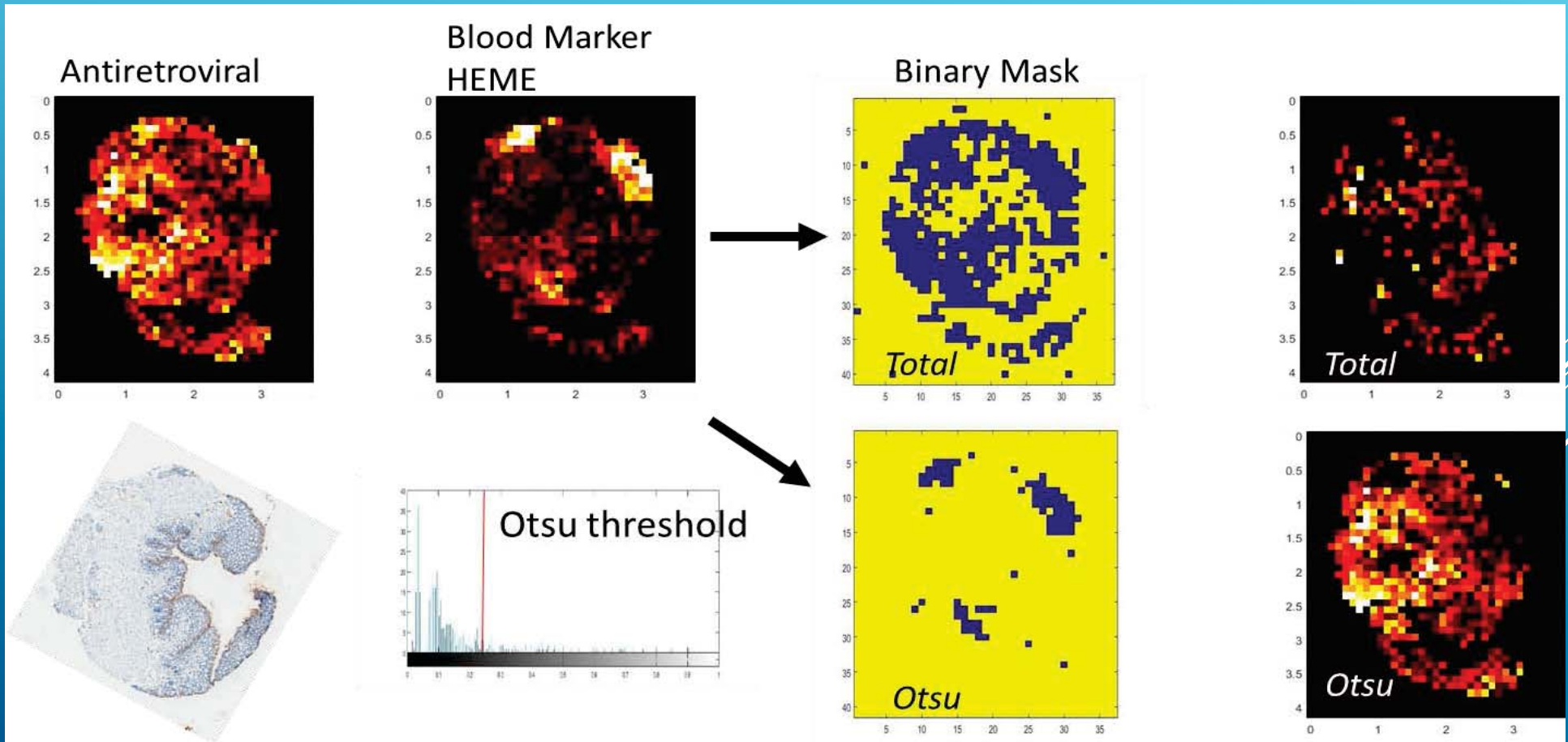
IR-MALDESI MSI



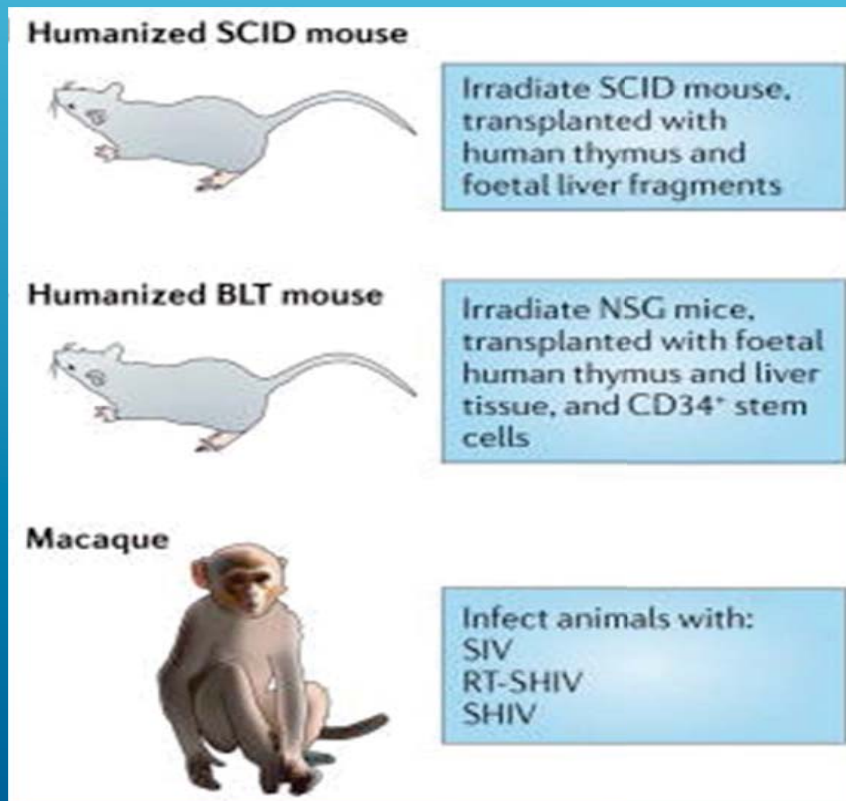
ISH/IHC



MSI OF ARVS IN TISSUES: *CORRECTING FOR BLOOD CONTAMINATION*



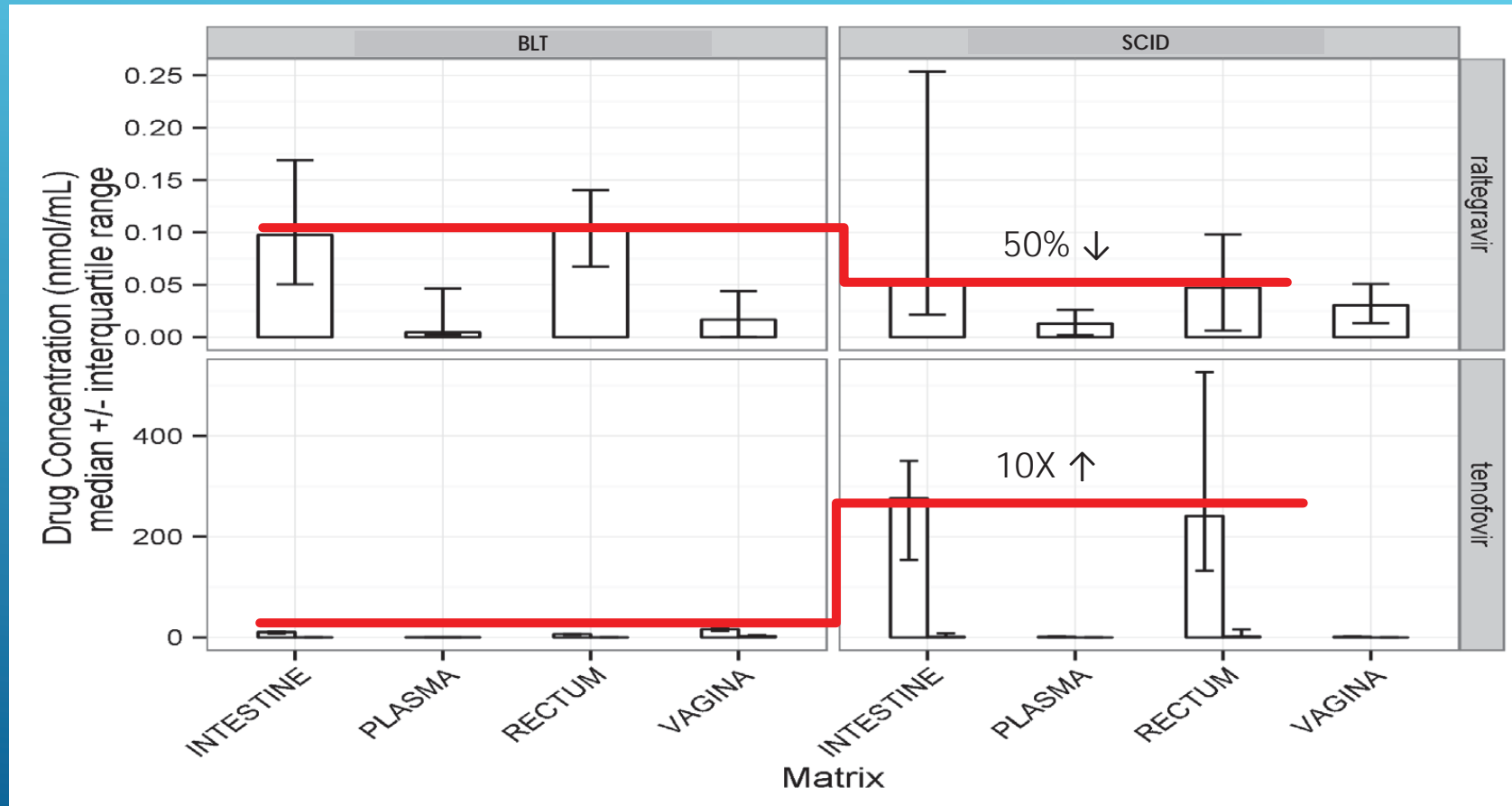
FACTORS AFFECTING DRUG DISTRIBUTION IN ANIMALS



- ▶ Drug distribution differences?
 - ▶ Drug transporter activity
 - ▶ Protein binding
 - ▶ Intracellular activation
 - ▶ Other local barriers
- ...changing the way drug is distributed from one model to the next

ANIMAL ARV TISSUE DISTRIBUTION VARIABLE BETWEEN MODELS

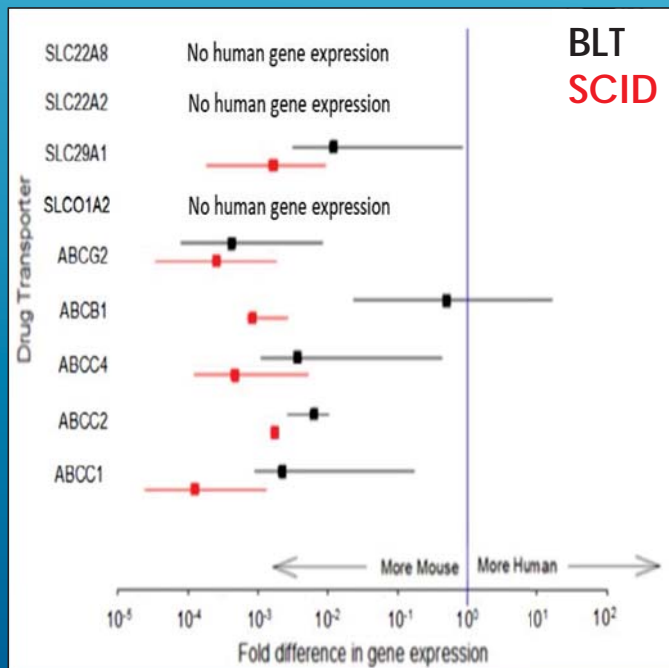
RALTEGRAVIR AND TENOFOVIR IN BLT VS SCID MICE GIVEN THE SAME DOSE



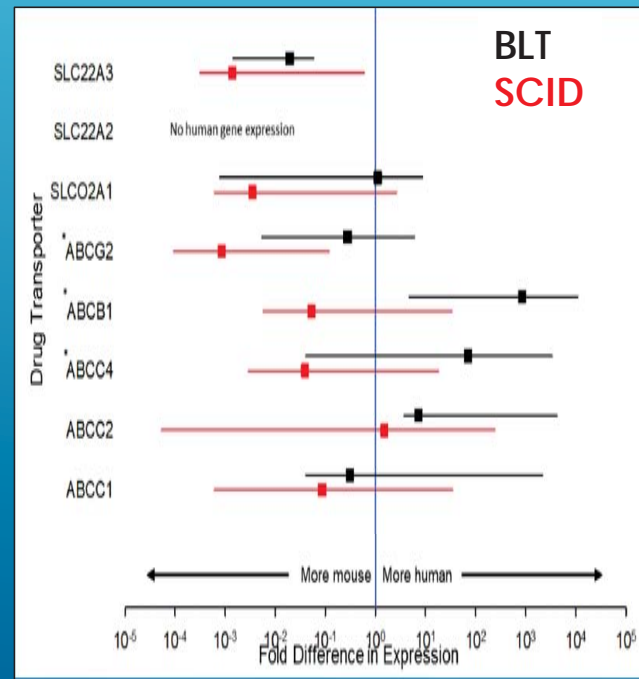
Denton et al. IAS 2013. Plos Pathogens 2013, Akkina et al IAS 2013

HUMANIZED MOUSE DIFFERENCES IN TRANSPORTER GENE EXPRESSION

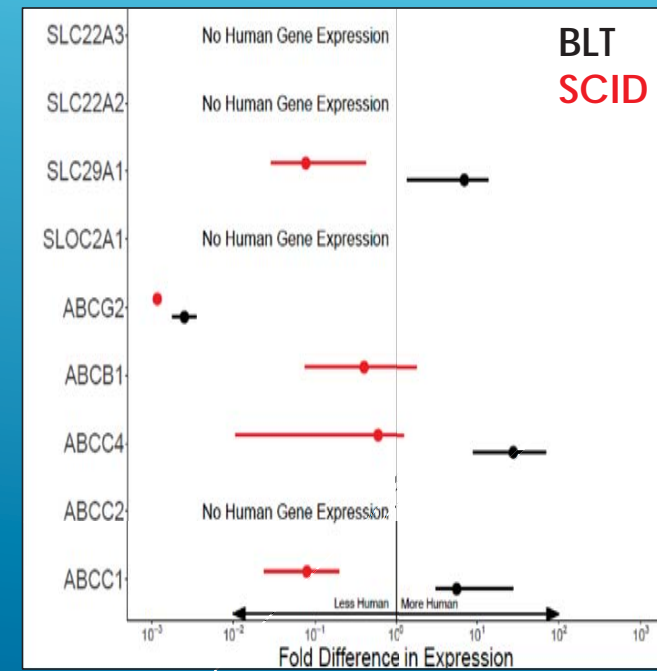
BRAIN



SPLEEN



ILEUM



SUMMARY

- ▶ HIV infection still a major cause of morbidity
 - ▶ Once ART is stopped, viral rebound inevitably occurs
- ▶ Long lived infected cells in privileged anatomic sites cause of slow reservoir decay
 - ▶ active viral replication in tissues is controversial, but likely occurs to some extent
- ▶ Cure strategies primarily focused on a functional cure
 - ▶ a combination of small molecules and immunotherapy (eg shock and kill)
- ▶ Unclear which latency reversal model (cells and animals) will be most predictive of efficacy

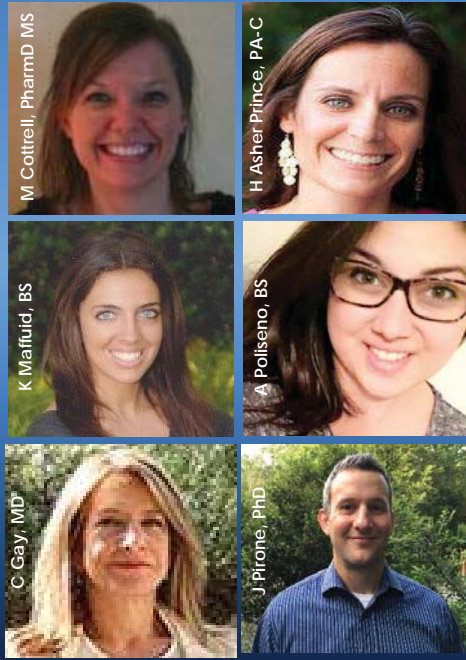
SUMMARY

- ▶ Pharmacologic insights will be critical to streamlining drug development
 - ▶ Hollow fiber cell models for PK/PD screening to identify dosing strategies, promising combination therapy, and optimal sequencing of combinations
 - ▶ Mass Spec Imaging promising for studying drug distribution and effect in tissues
 - ▶ Better prediction of drug penetration in tissue compartments
 - ▶ Understanding species differences in tissue PK/PD to develop accurate allometry for cure
 - ▶ If we cure a mouse/NHP can we cure a human?

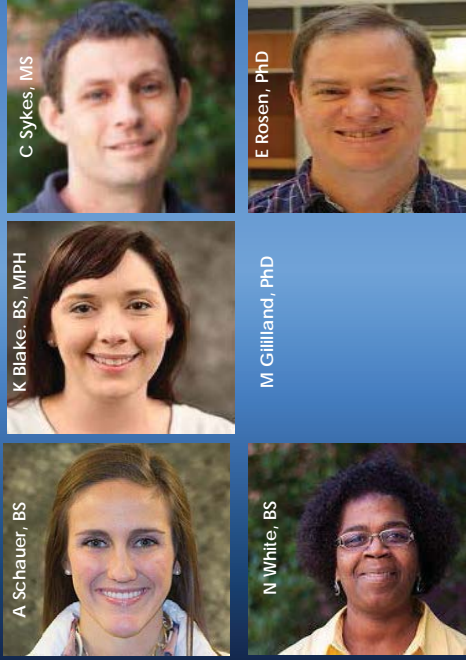
ACKNOWLEDGEMENTS

KASHUBA LAB MEMBERS

Clinical Pharmacology/ Pharmacometrics/ Clinical Trials



Bioanalytical Chemistry/ Mass Spectroscopy Imaging



Laboratory Operations/ QA



Learners

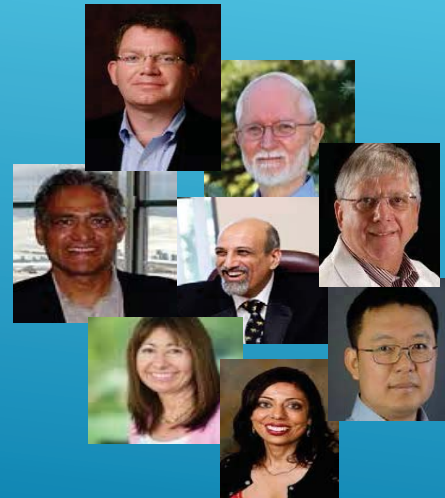


ACKNOWLEDGEMENTS

UNC Collaborators



External Colleagues/Collaborators



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 CONRAD

K23 AI093156
 T32 GM086330
 K23 HD064814
 R37 DK049381
 R56 AI091547
 K23 AI077355

U01 AI095031
 P30 AI050410
 U19
 AI1096113
 S10 RR026581
 P01
 MH094177
 R01 AI096138



IT ALWAYS SEEMS
IMPOSSIBLE
UNTIL IT'S DONE.

- NELSON MANDELA

LABORATORY HAZARD INFORMATION
1100 GMRB
EMERGENCY CONTACT INFORMATION
EMERGENCY: DIAL 911





LYMPHOID TISSUE MSI: *INTESTINAL TRANSPORTER OVERLAY*

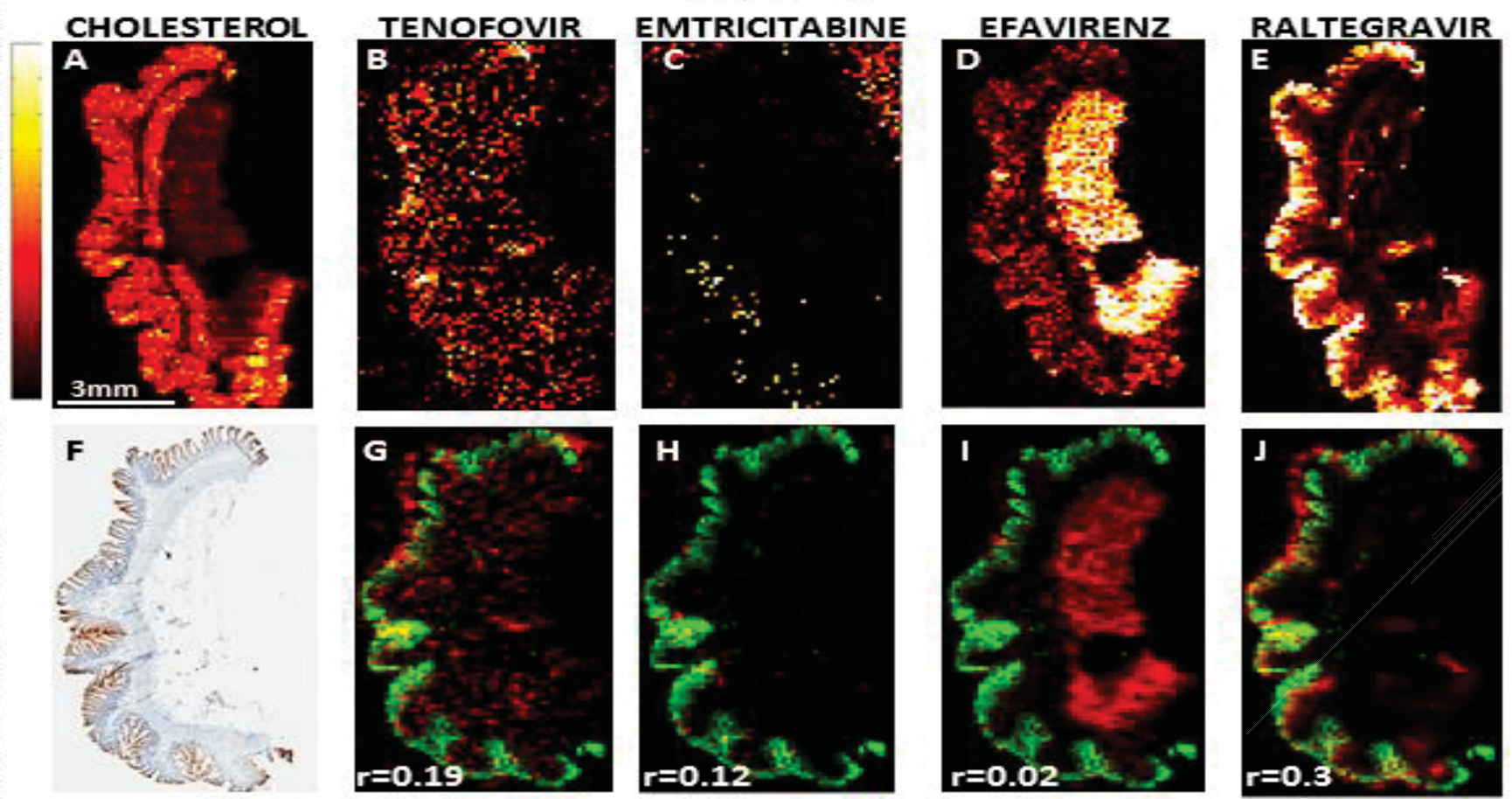
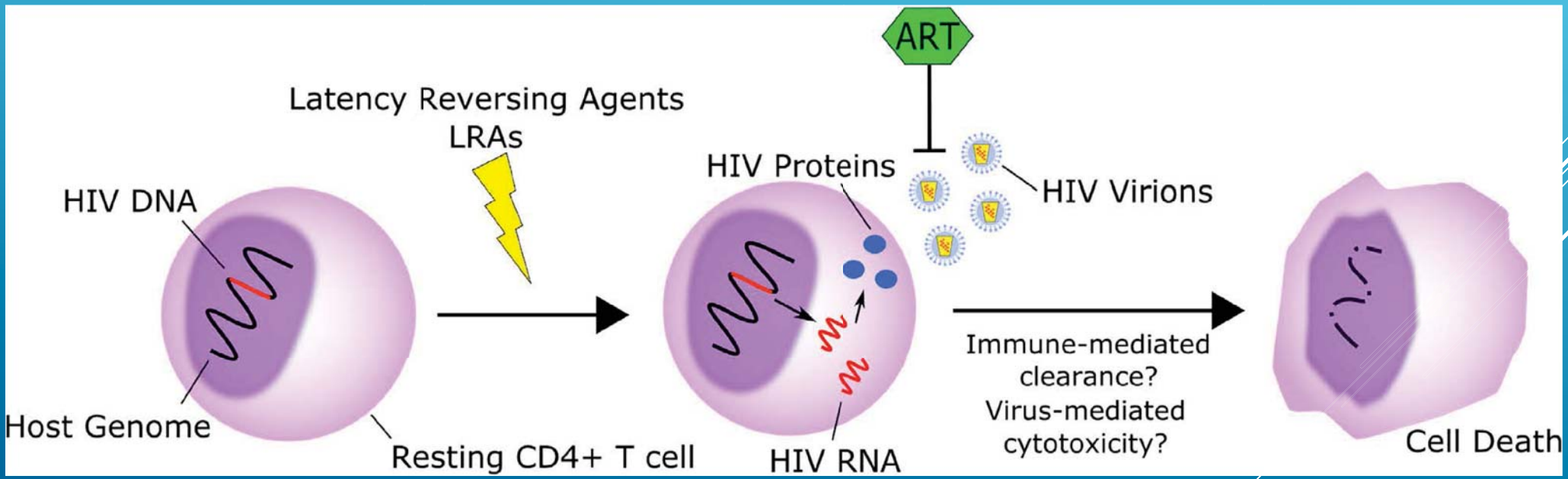


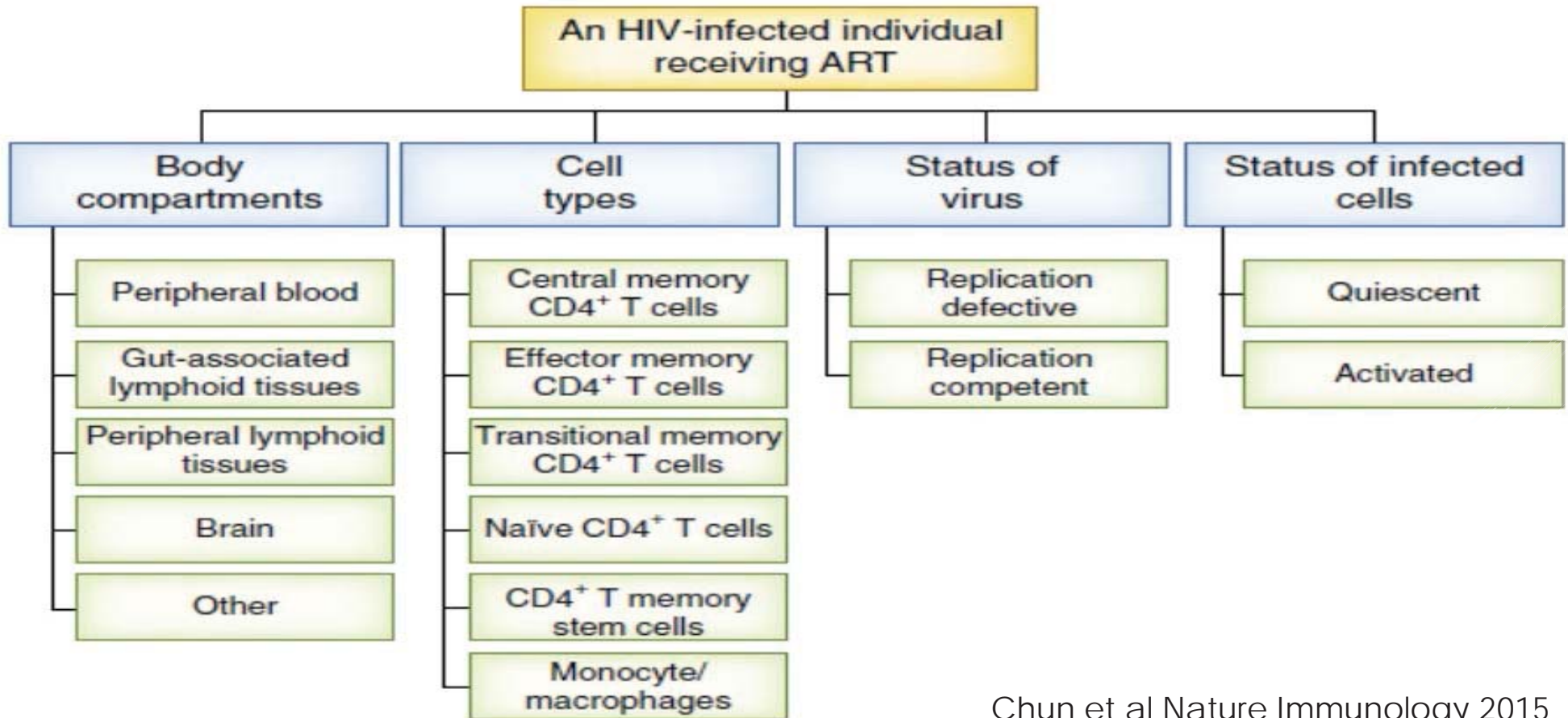
Table 1
Characteristics of the selected studies.

Author	HDAC inhibitor	Number of Participants	HDAC inhibitor's Regimen	Changes in Resting CD4+ T cells HIV RNA	Other outcomes
Chen et al. [28]	Vorinostat	5	Daily Vorinostat Monday through Wednesday for 8 weekly cycles	After dose 11 (second dose of cycle 4) or dose 22 (second dose of cycle 8) increased significantly in only 3 of the 5 participants, and the magnitude of the increase was much reduced compared with that after a single dose	Changes in histone acetylation were blunted. Quantitative viral outgrowth and total cellular HIV DNA were unchanged
Chen et al. [25]	Vorinostat	8	A single oral 200 mg dose for assessing tolerability, then 400 mg dose 4 (or more) weeks later	An increase of 1.5- to 10.0-fold (mean 4.8) in expression of unspliced HIV-1 gag RNA within resting CD4+ T cells was measured in seven patients	
Scott et al. [28]	Vorinostat	20	Vorinostat was administered 400 mg orally once daily for 14 days while maintaining ART	Cell associated unspliced HIV RNA in blood increased significantly in 18/20 patients (90%) with a median fold change from baseline to peak value of 7.4 (IQR 3.4, 9.1).	There were no statistically significant changes in plasma HIV RNA, concentration of HIV DNA, integrated DNA, inducible virus in CD4+ T-cells or markers of T-cell activation.
Smith et al. [35]	Romidepsin	20	Participants received 6 therapeutic intradermal HIV-1 immunizations with 12 mg/mL Vacc-4 x and 0.6 mg/mL rhuGM-CSF over 12 weeks (at 0 weeks, 1 week, 2 weeks, 3 weeks, 11 weeks, and 12 weeks) before receiving 5 mg/m ² intravenous Romidepsin once a week for 3 weeks.	No major changes in the CD4 T-cell compartment during Romidepsin infusions	Total HIV-1 DNA declined from screening to 6 weeks after Romidepsin treatment (mean reduction 39.7%, 95% CI -59.7 to -11.5; p = .012). The decrease in integrated HIV-1 DNA from baseline to 8 weeks after Romidepsin treatment was not significant between four (24%) and eight (47%) of 17 patients had detectable plasma HIV-1 RNA throughout the course of the study (19.2%, -38.6 to 6.3; p = .123).
Smits et al. [32]	Panobinostat	15	Oral Panobinostat 20 mg 3 times/week every other week for 8 weeks while maintaining combination antiretroviral therapy (cART)	Levels of CA-US RNA increased significantly during Panobinostat treatment (p < .0001) with significant increases on time points on-treatment as compared to baseline. The median maximal fold-increase in CA-US RNA was 3.5 (range 2.1-14.4). Levels of CA-US RNA remained elevated 4 weeks post-Panobinostat (fold-increase 1.60; 95% CI: 1.17-2.19; P = .003)	Using a transcription mediated amplification-based semi-quantitative assay (Procleix Ultrio Plus, 59% analytic sensitivity of 3.6 copies/mL), HIV-RNA in plasma was detected more frequently during Panobinostat administration with an odds ratio of 10.5 (95% CI: 2.2-50.3) for a positive test on-treatment compared to baseline
Wang et al. [29]	Romidepsin	6	One 4 h Romidepsin infusion (5 mg/m ²) per week for three consecutive weeks and were followed for up to 70 days after the last infusion	HIV-1 transcription quantified as copies of cell-associated un-spliced HIV-1 RNA increased significantly from baseline during treatment (range of fold-increase: 2.4-5.0; p = .03).	Plasma HIV-1 RNA increased from < 20 copies/mL at baseline to readily quantifiable levels at multiple post-infusion time-points in 5 of 6 patients (range 46-103 copies/mL following the second infusion, p = .04).
Wang et al. [37]	Romidepsin	20	Six Vacc-4 x (1.2 mg) intradermal immunizations using rhuGM-CSF (60 µg) as adjuvant were followed by 3 weekly intravenous infusions of romidepsin (5 mg/m ²).		Participants with CD8+ T-cell proliferation assay positivity post-vaccination showed reductions in total HIV DNA post-vaccination (p = .006; q = 0.183), post-latency reversal (p = .005; q = 0.183), and CA-RNA reductions post-vaccination (p = .015; q = 0.254). Participants (40%) were defined as proliferation 'Responders' having ≥2-fold increase in assay positivity post-baseline. Robust ELISpot baseline responses were found in 87.5% participants. No significant changes were observed in the proportion of polyfunctional CD8+ T-cells to HIVGag by ICS. There was a trend towards increased viral inhibition from baseline to

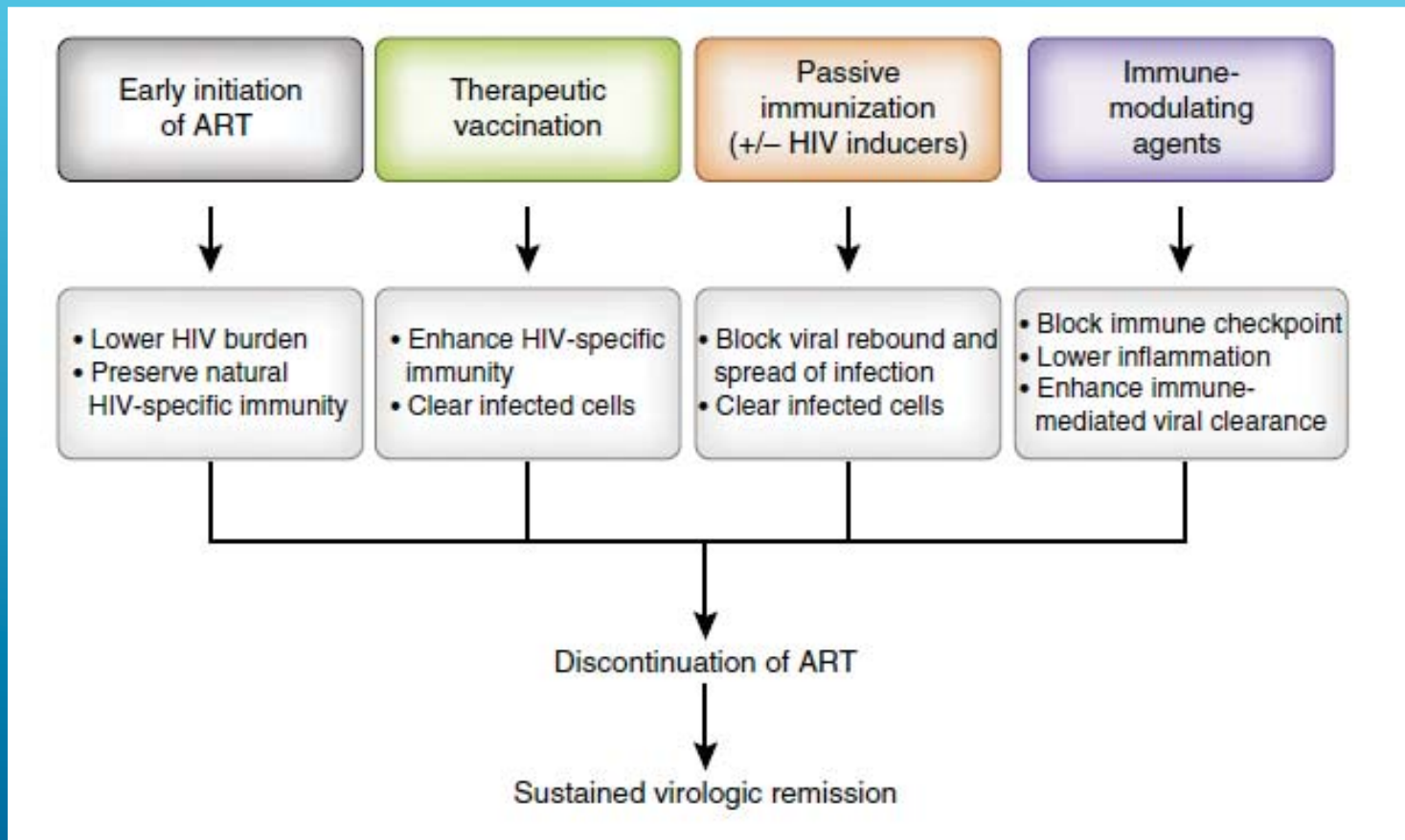


Kim et al. Cell Host and Microbe, 2018

COMPLEX NATURE OF HIV RESERVOIRS



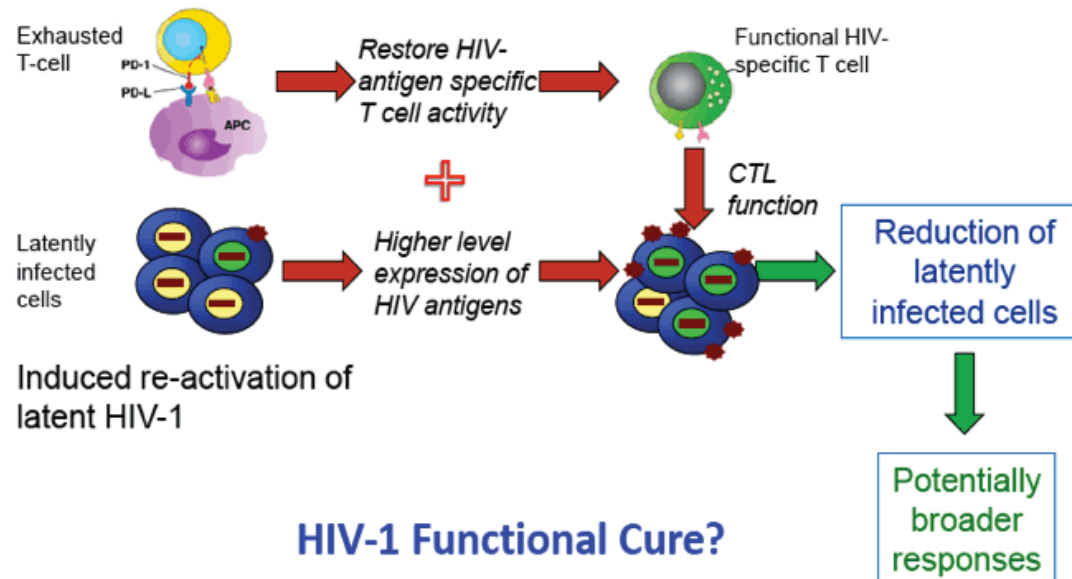
Chun et al Nature Immunology 2015

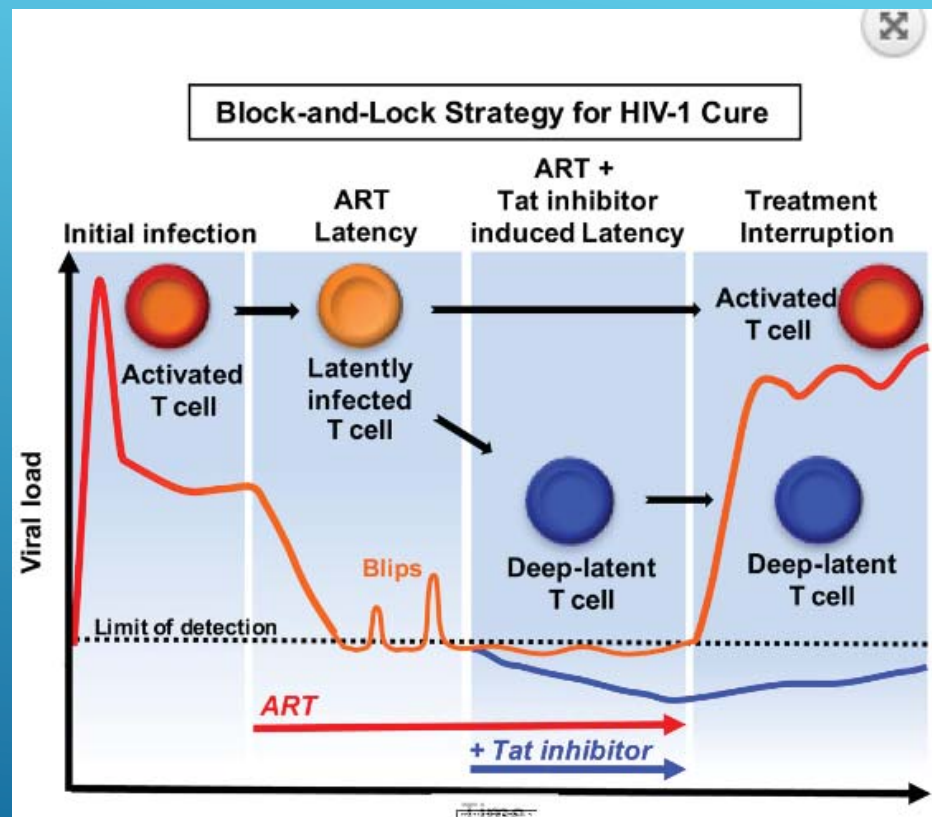


Chun et al Nature Immunology 2015

BMS Strategy for HIV-1 Functional Cure: *Dual Approach*

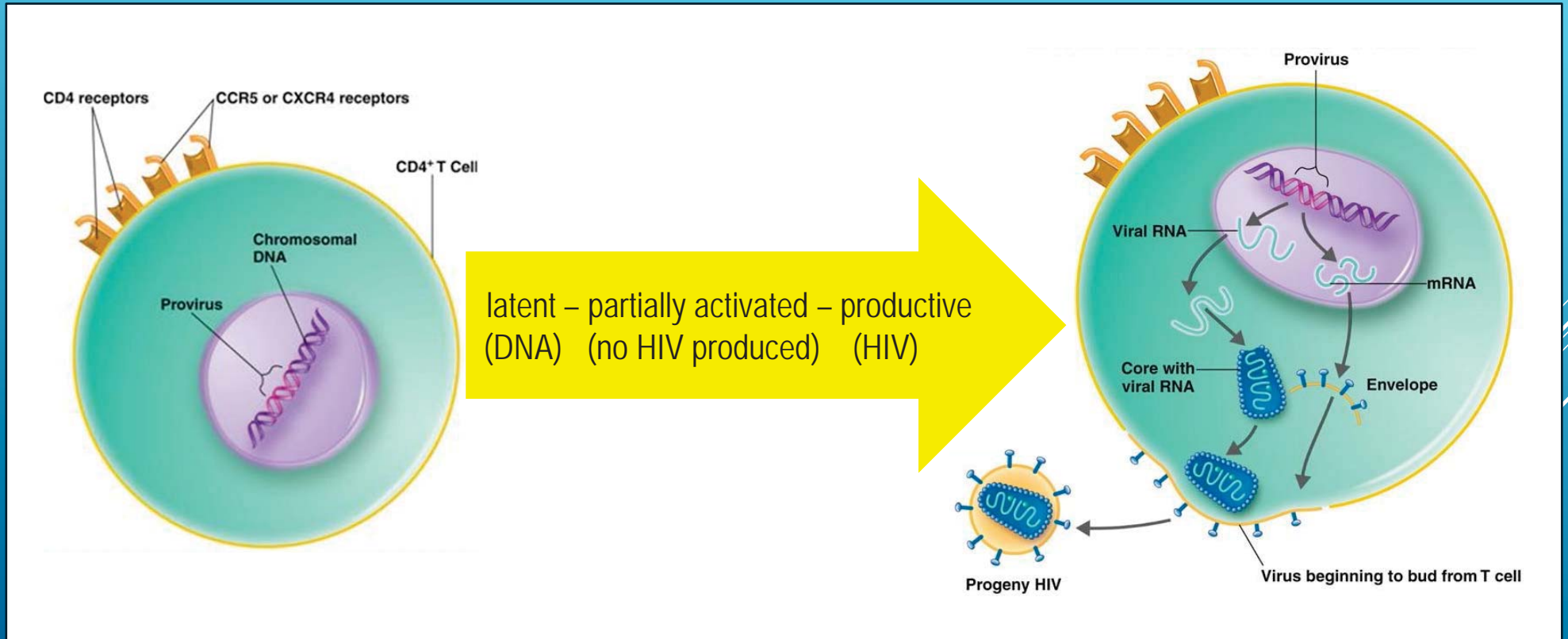
Treatment with α PD-L1





re (Kessing et al., 2017; doi:
10.1016/j.celrep.2017.09.080)

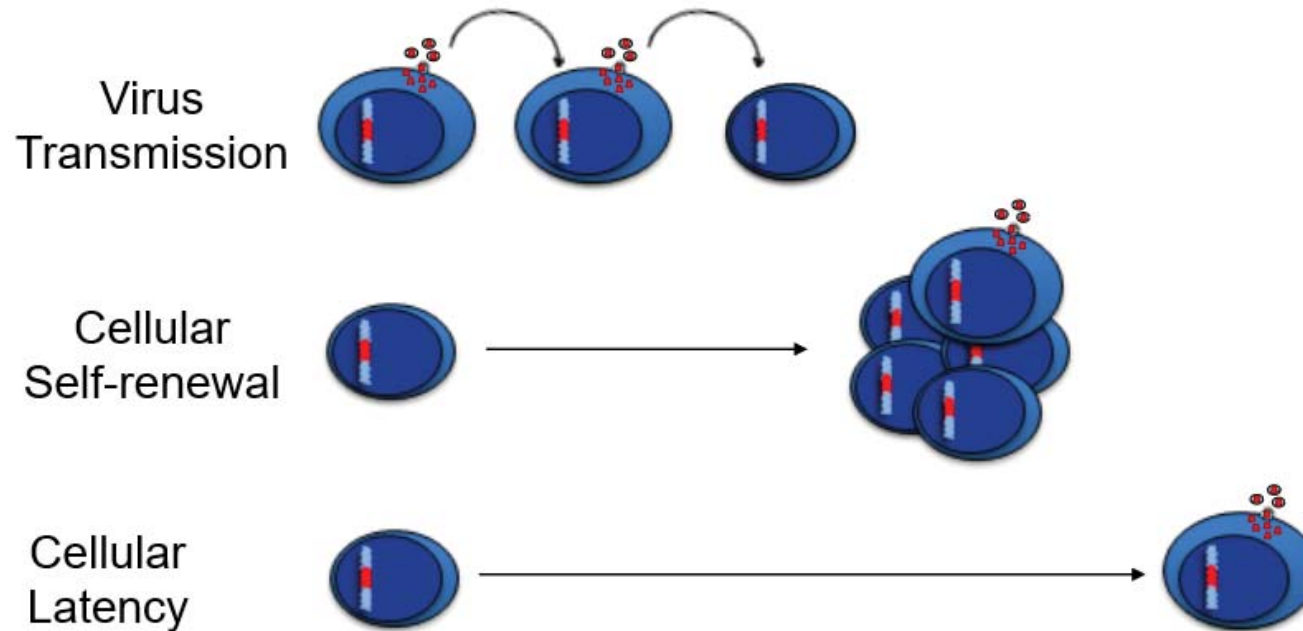
SUBCLASSES OF INFECTED CELLS



Fischer, Retrovirology 2008
Althaus, PLoS Comp Biol, 2015
Yuki, STM, 2018

Adapted from Huldrych, CROI 2018

Three Histories of HIV-infected Cells In Vivo



Multiple mechanisms govern these events

Boston Patients: Virus Recrudescence

Hematopoietic transplantation with cells susceptible to infection

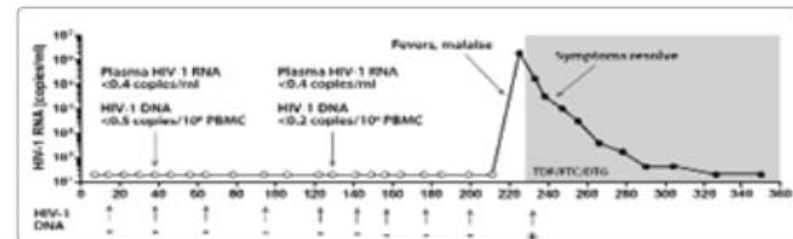
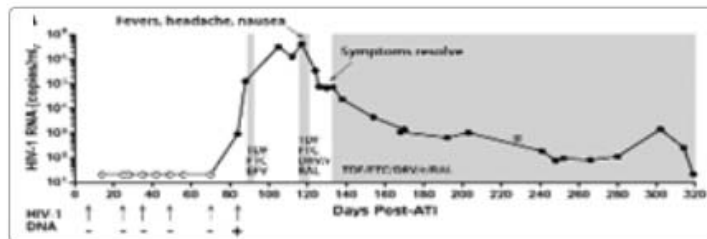
Annals of Internal Medicine

ORIGINAL RESEARCH

Antiretroviral-Free HIV-1 Remission and Viral Rebound After Allogeneic Stem Cell Transplantation

Report of 2 Cases

Timothy J. Henrich, MD; Emily Hanauer, BS; Francisco M. Marty, MD; Michael N. Strigam, BS; Sheila Kwock, PhD; Tsung-Hua Lee, MD, PhD; Yvonne P. Robbins, BS; Benjamin T. Davis, MD; Jonathan Z. Li, MD; Andrea Heston, BS; Alison L. Hill, PhD; Michael P. Busch, MD, PhD; Philippe Armand, MD, PhD; Robert J. Soffler, MD; Marcus Altmann, MD, PhD; and Daniel R. Kuritzin, MD



Despite 1000 – 10,000 fold reductions in reservoir size, virus rebounded
Modeling: latent reservoir will have to be depleted $> 10^5$ fold (Hill, *PNAS* '14)

A single virus accounts for recrudescence

From DC Douek, MD, MRCP, PhD at Atlanta, GA: April 8, 2016, IAS-USA.

Slide 11 of 29

STRATEGIES TO A CURE

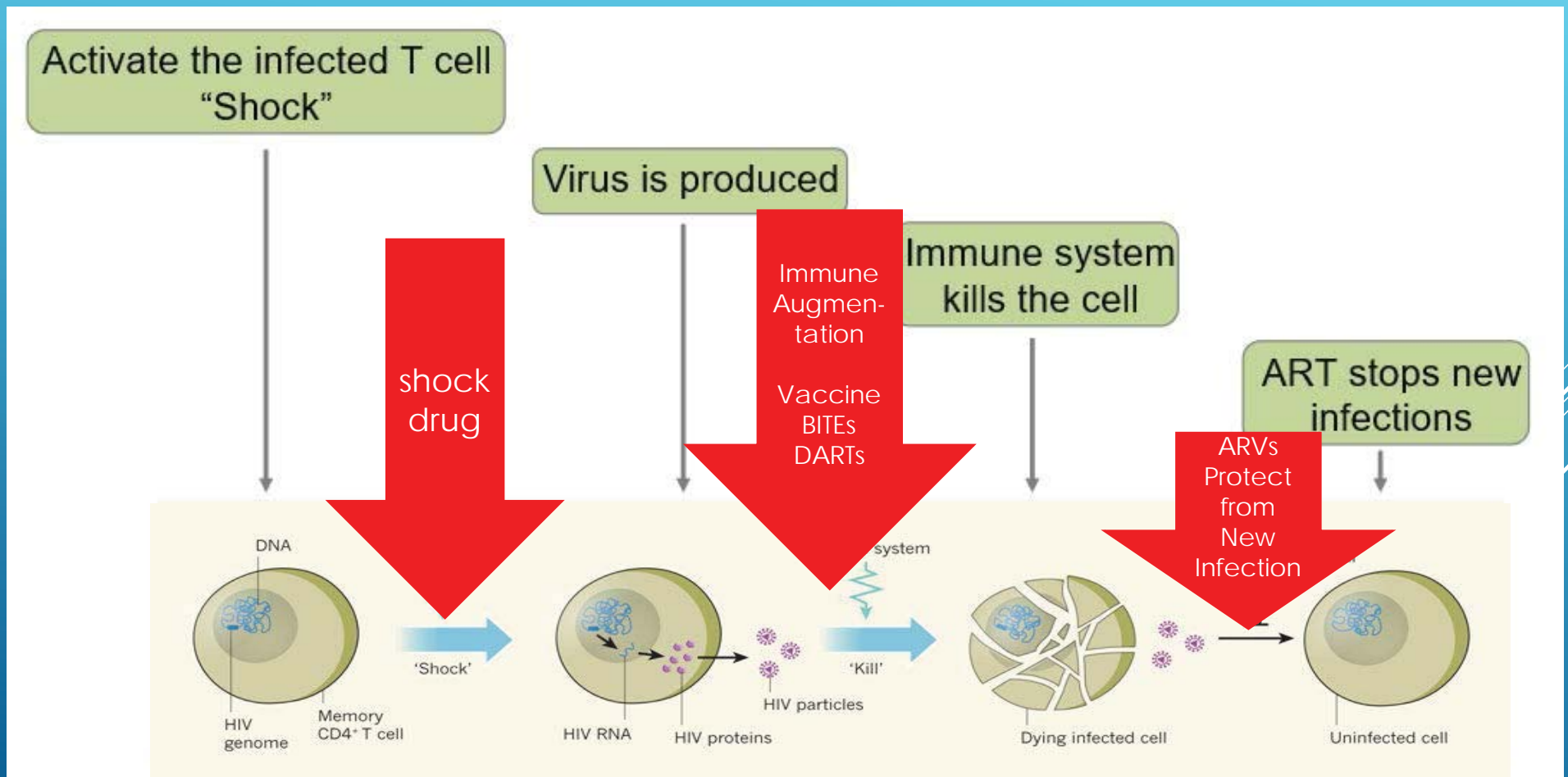
FUNCTIONAL CURE

- Immune mediated control of viral replication
- Eg elite controllers; boosting the immune system
- ART may be withdrawn without subsequent viral rebound (eg “complete remission” after treatment for malignant cancer)
- But these strategies may result in increased inflammation/ comorbidities

STERILIZING CURE

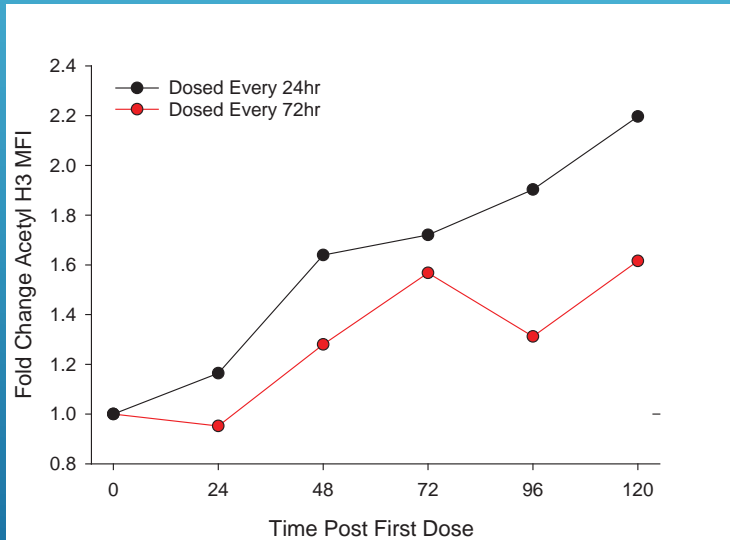
- Elimination of all HIV-infected cells from the body
 - infected cells not visible to host defenses
 - frequency of HIV-specific CD8 T cells typically decreases with ART and often have “exhausted” (or dysfunctional) phenotype
 - Infected cells broadly distributed to sites relatively inaccessible to host defenses or treatment

SHOCK AND KILL STRATEGY

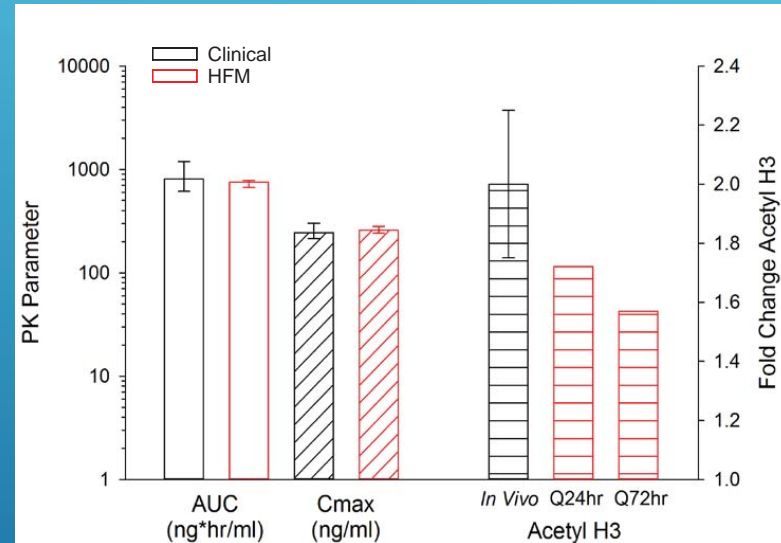


PREDICT PD BY SIMULATING *IN VIVO* PK

PD Measured over Time



Clinical vs *In Vitro* PK/PD



MSI (IR MALDESI)

QUANTITATIVE INFRARED MATRIX ASSISTED LASER DESORPTION ELECTROSPRAY IONIZATION

